

LC-HRMS-based Multi-Attribute Method for Oligonucleotides (MAMO): Characterization and Impurity Profiling

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The authors declare no competing financial interest.

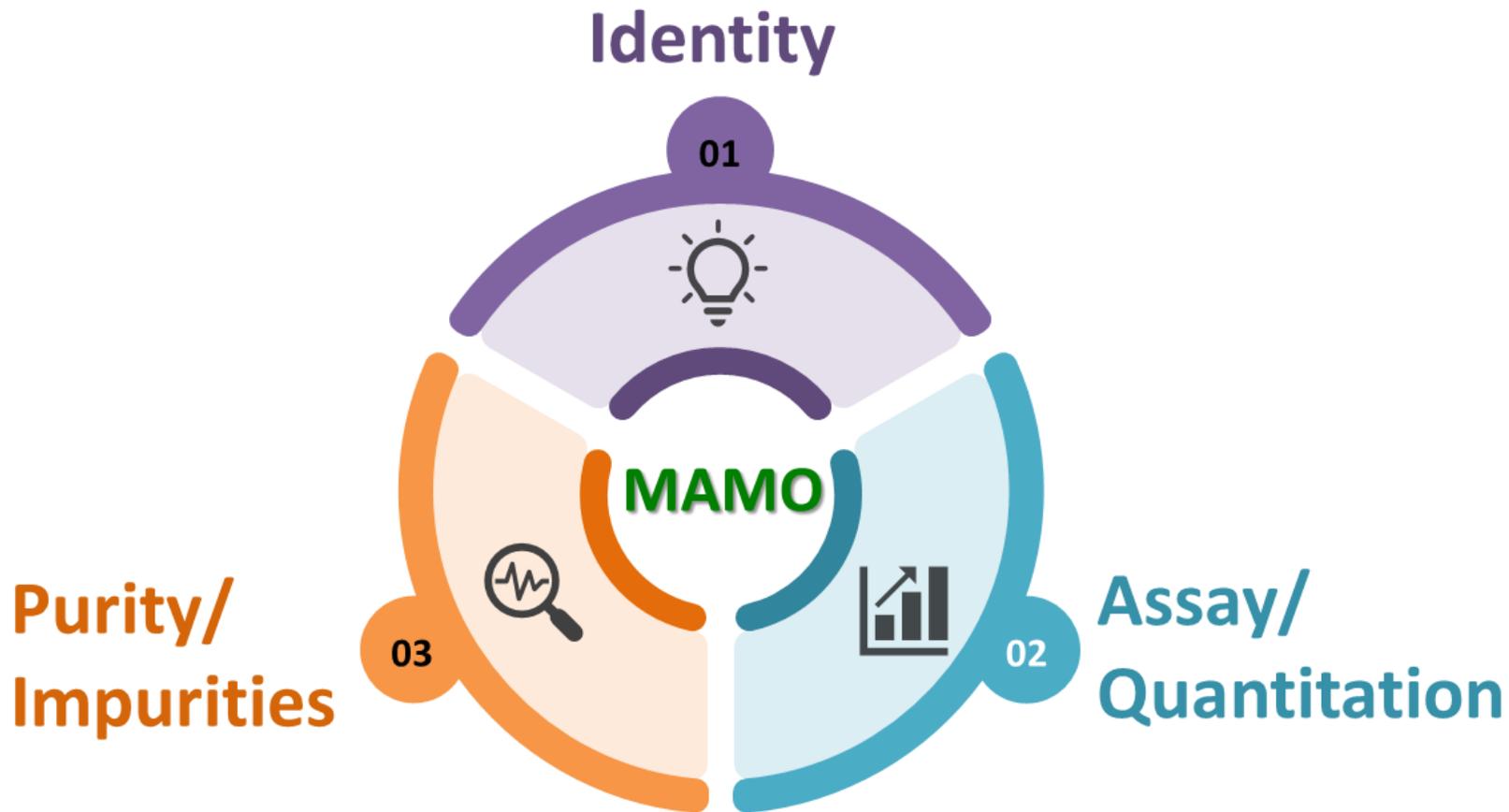
Every patient deserves confidence in their *every* and *next* dose of medicine.

Pharmaceutical quality assures the availability, safety and efficacy of *every* dose.

Outline

- LC-HRMS-based **Multi-Attribute Method** for **Oligonucleotides** (MAMO) analytical platform
- LC-HRMS method validation
- When separation by LC or MS fails

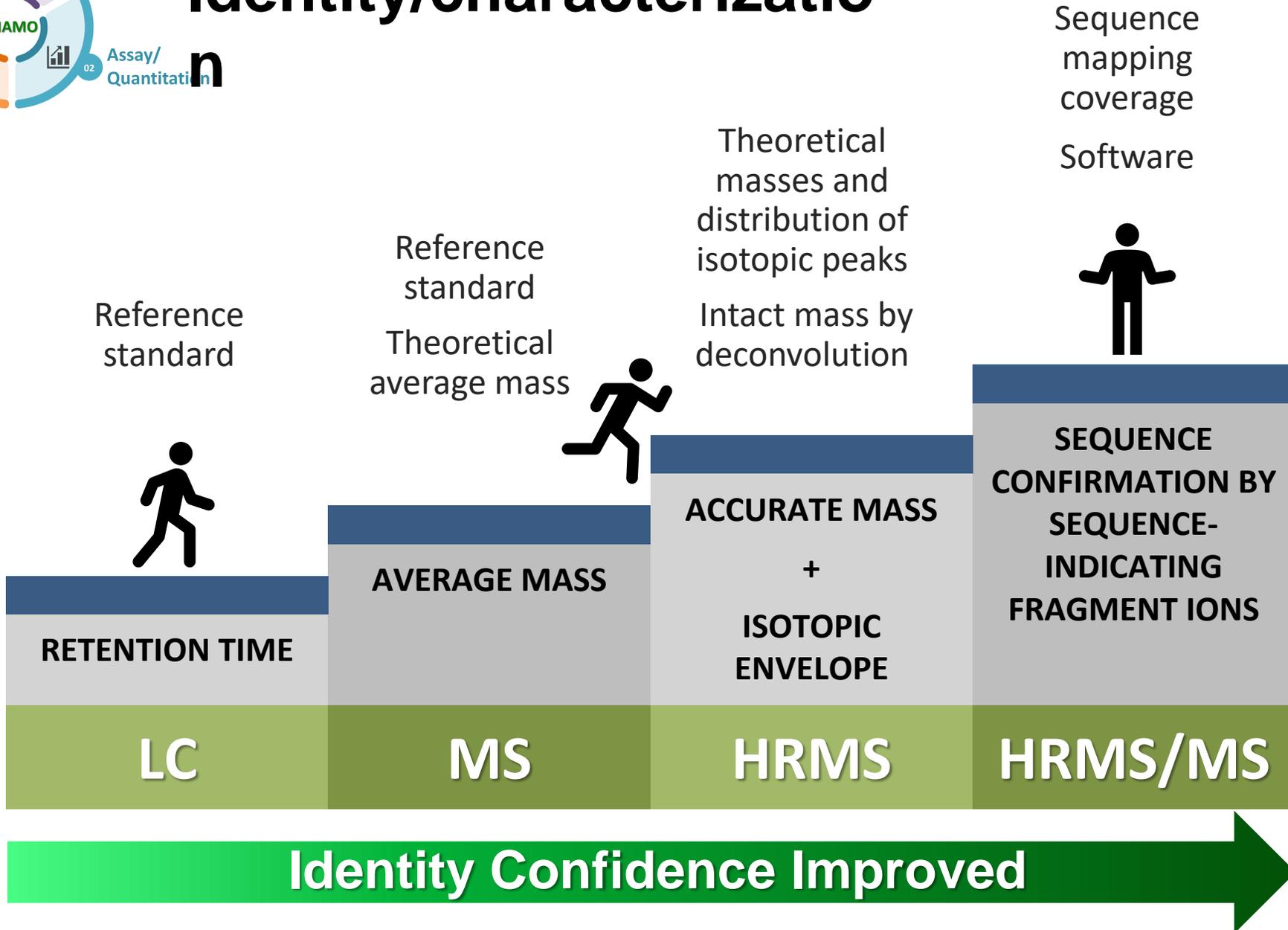
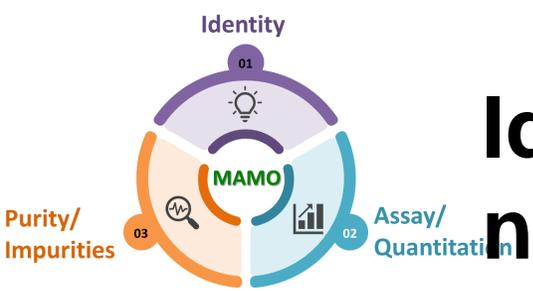
MAMO Analytical Platform



LC-HRMS-based

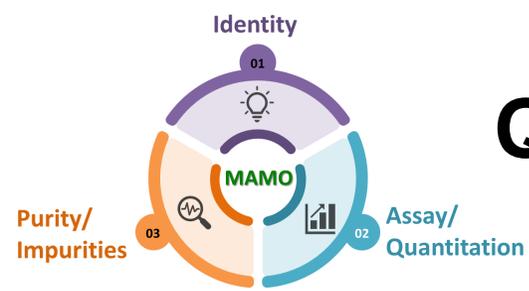
MAMO Method for Oligonucleotides

Identity/characterization



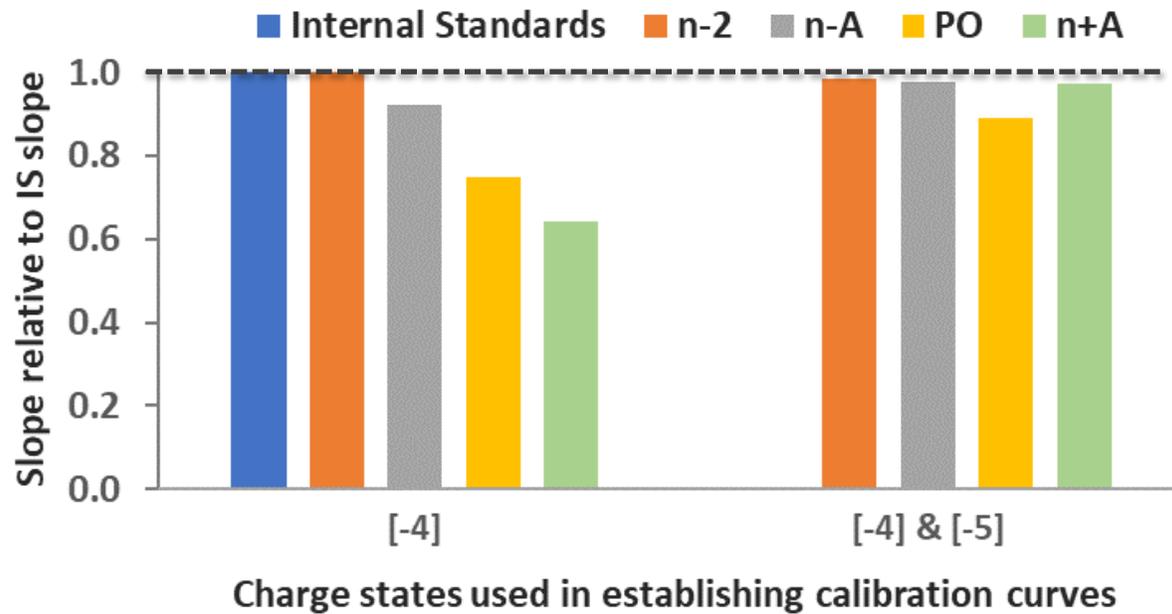
Abdullah, AM; Sommers, C.; Hawes, J.; Rodriguez J.; Yang, K. Tandem Mass Spectrometric Sequence Characterization of Synthetic Thymidine-rich Oligonucleotides. Journal of Mass Spectrometry. 2022, 57 (4), e4819

Quantification: Assay, Purity, Impurities



➤ Ionization efficiency

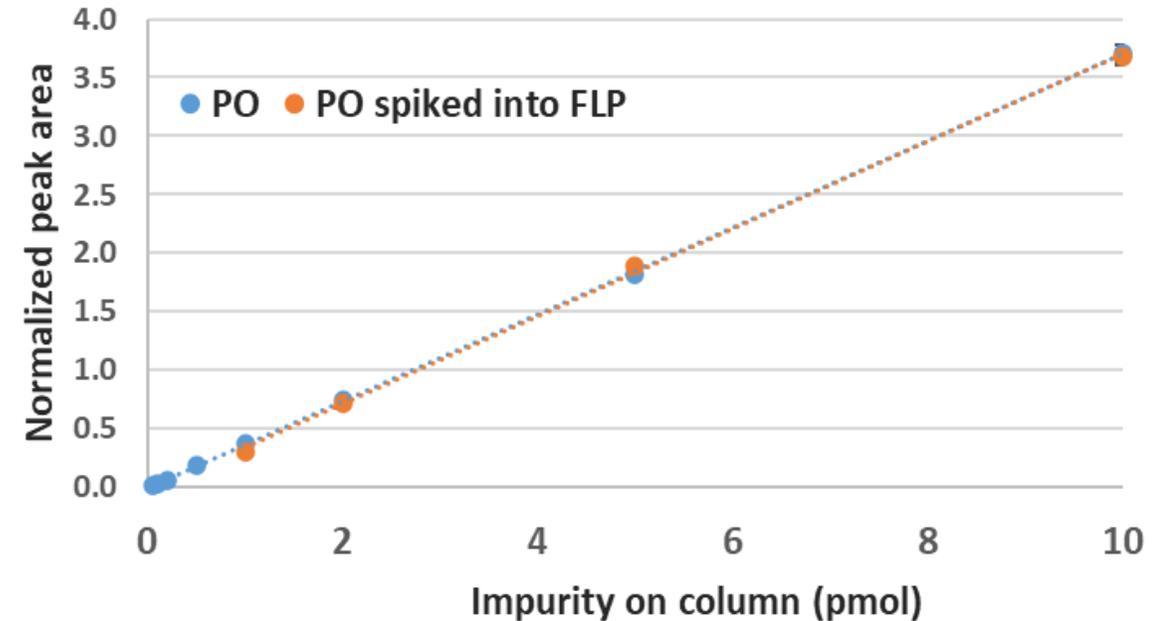
Slopes of calibration curves of impurities



n-2, n-A, 1PO, n+A: common impurities of nusinersen (18-mer)
 ISS: U₁₅, A₁₅, C₁₅

➤ Ion suppression

Calibration curves in the presence vs absence of high abundance FLP



FLP: full-length product (nusinersen sequence)

LC-HRMS Method Validation



✓ Hydrophilic interaction liquid chromatography (HILIC)- HRMS

System suitability testing

Specificity

Linearity

Range

Precision

Accuracy

LLOQ

Robustness

Excipient



Q2(R2) Validation of Analytical Procedures Guidance for Industry

U.S. Department of Health and Human Services
Food and Drug Administration
Center for Drug Evaluation and Research (CDER)
Center for Biologics Evaluation and Research (CBER)

March 2024
ICH-Quality
Revision 2

M10 BIOANALYTICAL METHOD VALIDATION AND STUDY SAMPLE ANALYSIS

Guidance for Industry

U.S. Department of Health and Human Services
Food and Drug Administration
Center for Drug Evaluation and Research (CDER)
Center for Biologics Evaluation and Research (CBER)

November 2022
ICH



System suitability testing



- Sensitivity check
 - LLOQ (peak area % CV)
 - Before and after sample assay
- Precision
 - Injector performance (peak area % CV)
 - Pump performance (retention time % CV)
 - Column performance (peak symmetry factor)
- Mass accuracy compared to theoretical value
 - ppm
- Linearity of injection accuracy and MS response
 - R^2
 - Before and after sample assay



Method validation

Specificity, Linearity, Range, **Precision**, Accuracy, LLOQ, Robustness, Excipient

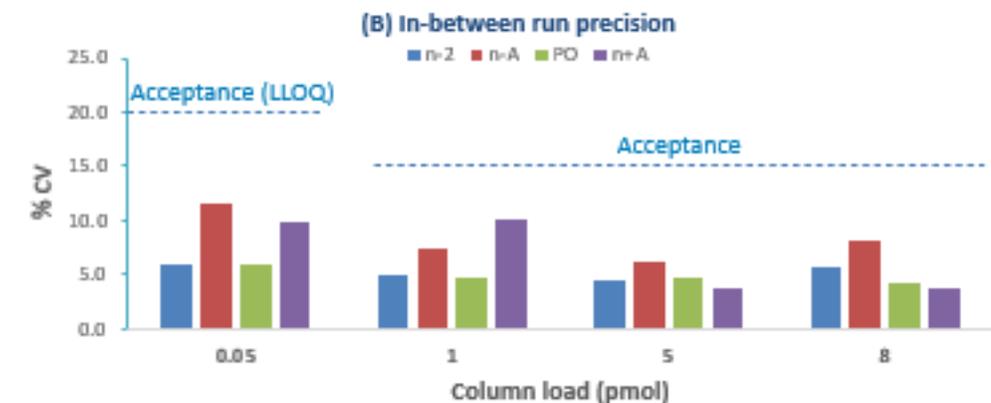
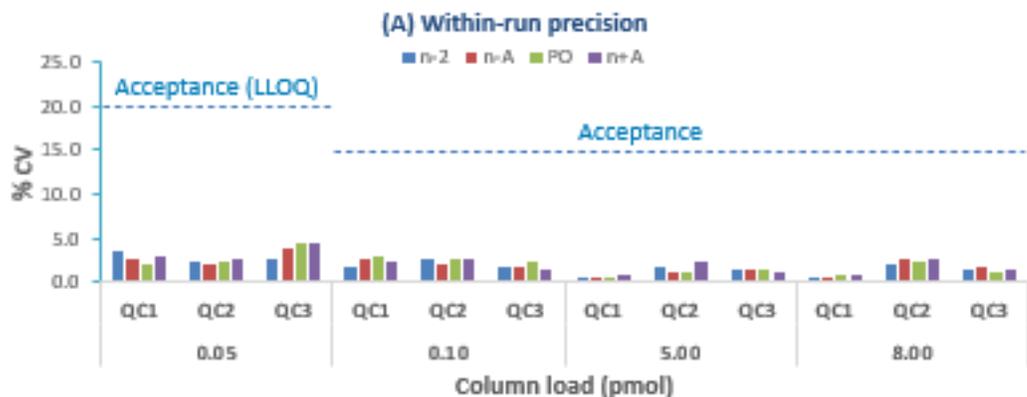


Fig. Precision (% CV) evaluated using 4 QC levels (0.05, 0.1, 5 and 8 pmol) run 3 different days (QC1, QC2 and QC3). (A) Within-run; and (B) In-between run

Table. % Recovery for three QC runs (QC1, QC2 and QC3) each comprising 4 QC levels

(A) % Recovery for n-2				(B) % Recovery for n-A			
Column load (pmol)	QC1	QC2	QC3	Column load (pmol)	QC1	QC2	QC3
0.05	93.69	102.37	94.42	0.05	86.57	104.40	87.61
0.10	96.87	99.33	99.87	0.10	94.20	100.68	103.79
5.00	104.47	98.67	100.53	5.00	98.91	101.80	99.66
8.00	101.74	101.25	97.91	8.00	95.97	104.29	96.86

(C) % Recovery for PO				(D) % Recovery for n+A			
Column load (pmol)	QC1	QC2	QC3	Column load (pmol)	QC1	QC2	QC3
0.05	89.31	104.77	95.55	0.05	86.40	101.06	97.08
0.10	88.34	97.00	96.97	0.10	81.37	96.19	105.34
5.00	99.24	94.92	104.31	5.00	100.92	97.40	108.96
8.00	95.95	97.08	100.79	8.00	97.86	98.77	105.70

Product-related Impurities

Deletion/Addition Sequences

$n-1, n-2$
 $n+1, n+2$
...

$n-G, n-A,$
 $n-U/n-C$
...

$n-G_1, n-G_2, \dots$
 $n-A_1, n-A_2, \dots$
... ..

Modified Backbone,
Nucleobases,
Sugar;
Cross-linking
...

Classes

Families

Members

Components

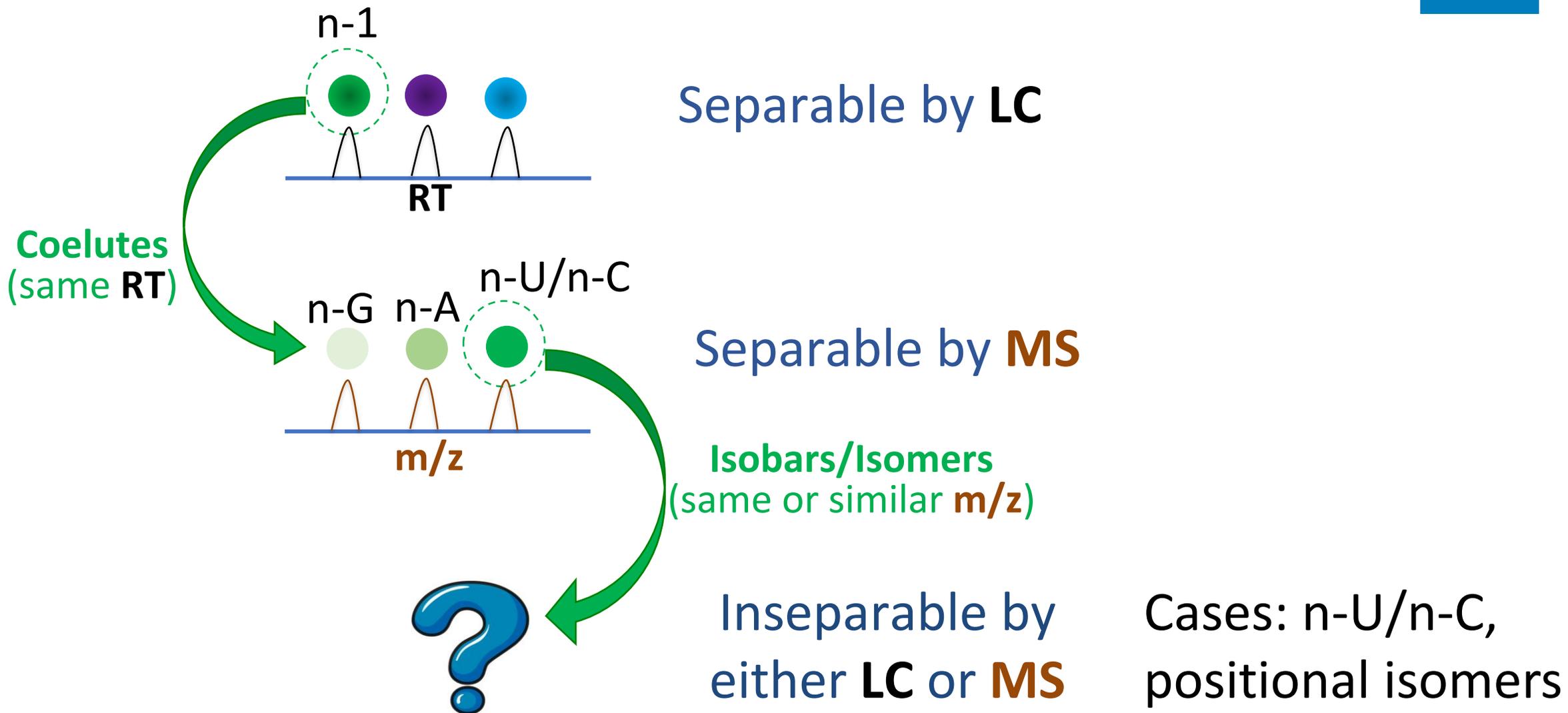
Tier-1:
Chromatographically separated

Tier-2:
Coeluting,
Different masses

Tier-3:
Coeluting,
Identical mass

Analytical challenge
↓
High

When Separation by LC or MS Fails



? Coeluting isobaric impurity ions



analytical
chemistry

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Article

Decoding Complexity in Synthetic Oligonucleotides: Unraveling Coeluting Isobaric Impurity Ions by High Resolution Mass Spectrometry

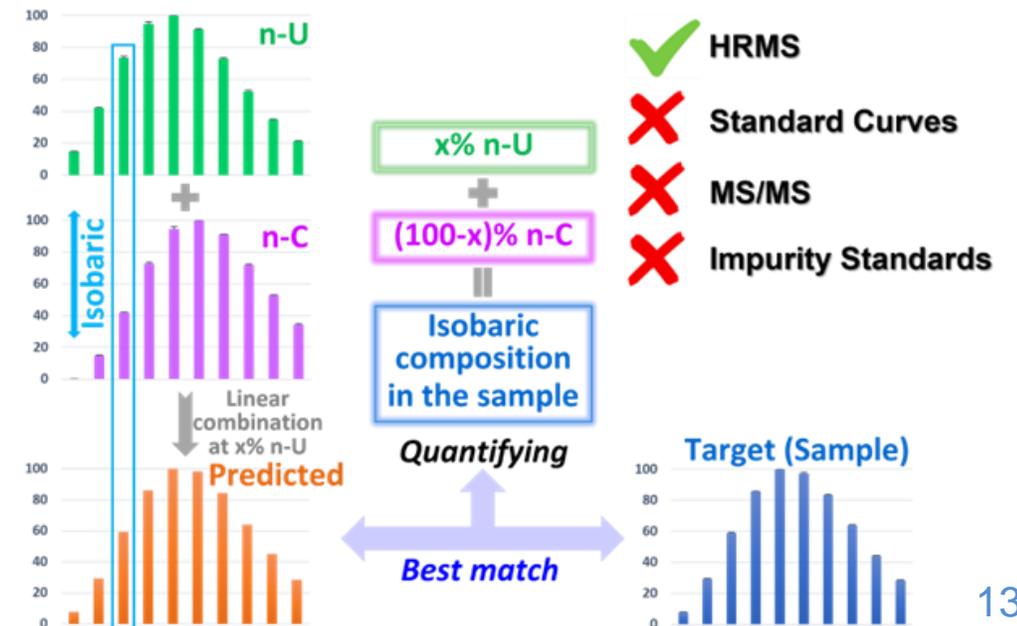
A. M. Abdullah, Cynthia Sommers, Jason D. Rodriguez, Deyi Zhang, Darby Kozak, Jessica Hawes, Mohan Sapru, and Kui Yang*

Cite This: *Anal. Chem.* 2024, 96, 904–909

Read Online

Nusinersen sequence:

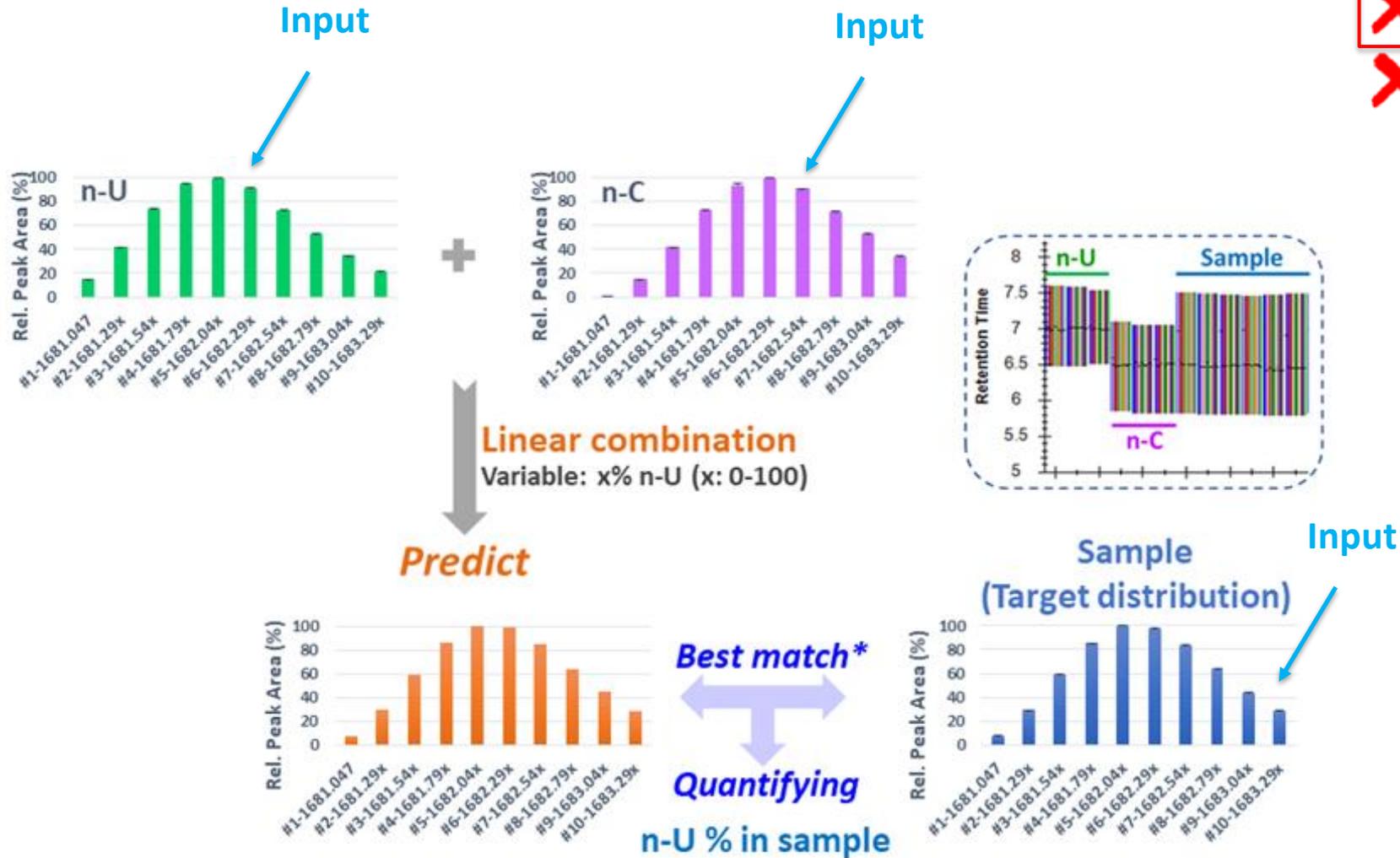
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? Coeluting isobaric impurity ions



- ✓ HRMS
- ✗ Standard Curves
- ✗ MS/MS
- ✗ Impurity Standards



Workflow

? Coeluting isobaric impurity ions

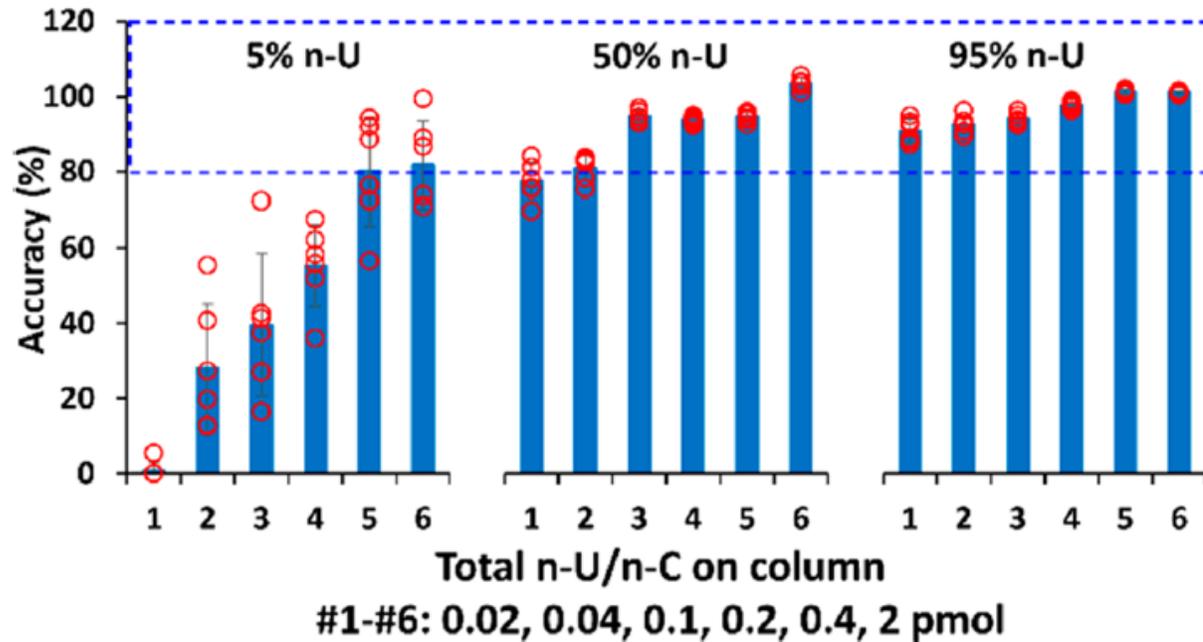


Figure 3. Quantified isobaric compositions of n-U/n-C mixtures at varied concentrations. The bar represents the mean and standard deviation (SD) values of replicate data points (red circles). The frame represents the same as that in Figure 2.

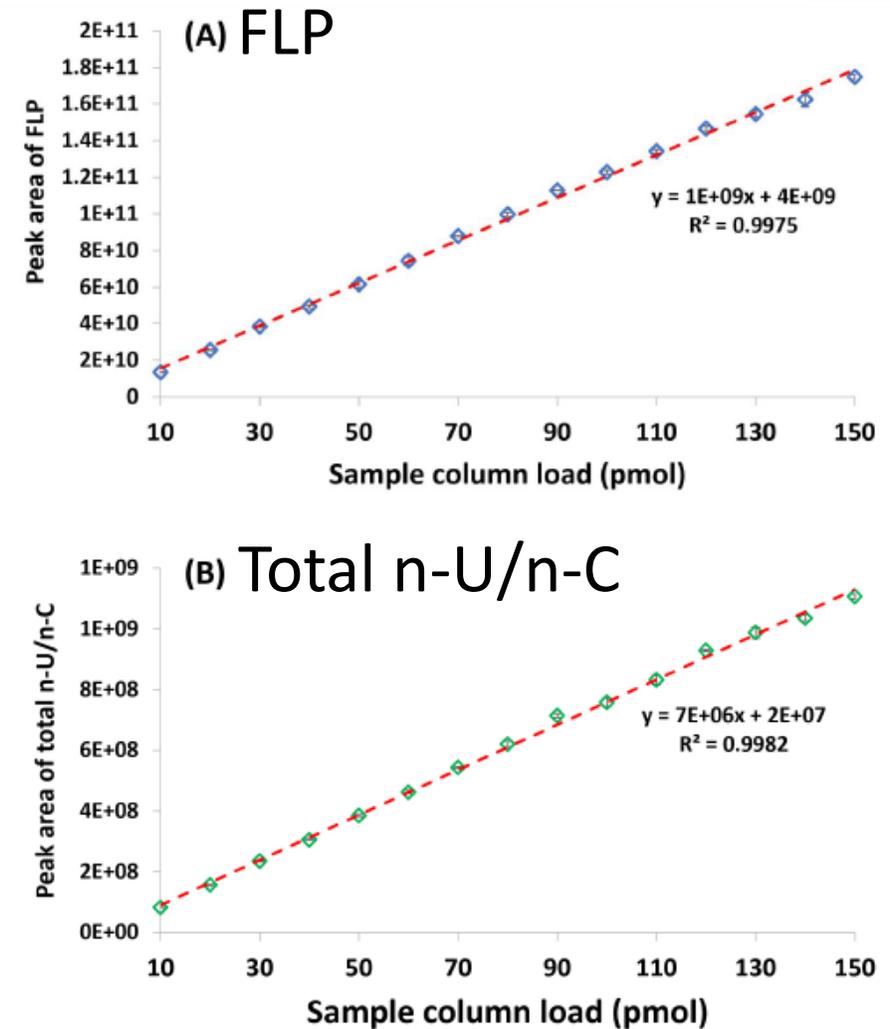


Figure S5. Linear responses in quantitation for FLP and total n-U/n-C impurity within the tested range of sample column loads ranging from 10 to 150 pmol. Quantitation was performed using full MS by total EIC peak area of 10 isotopic peaks for FLP (A) and n-U/n-C (B). Dashed line represents the linear regression.

? Coeluting isobaric impurity ions

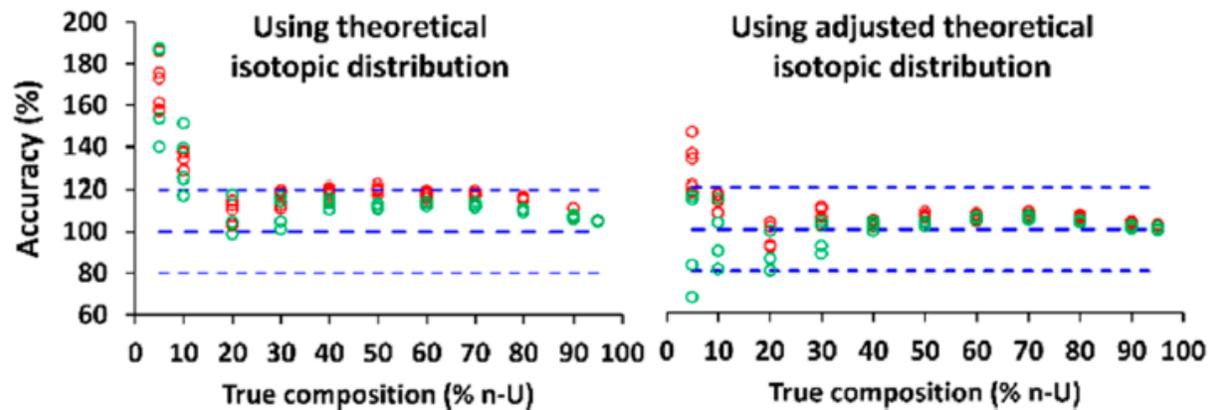
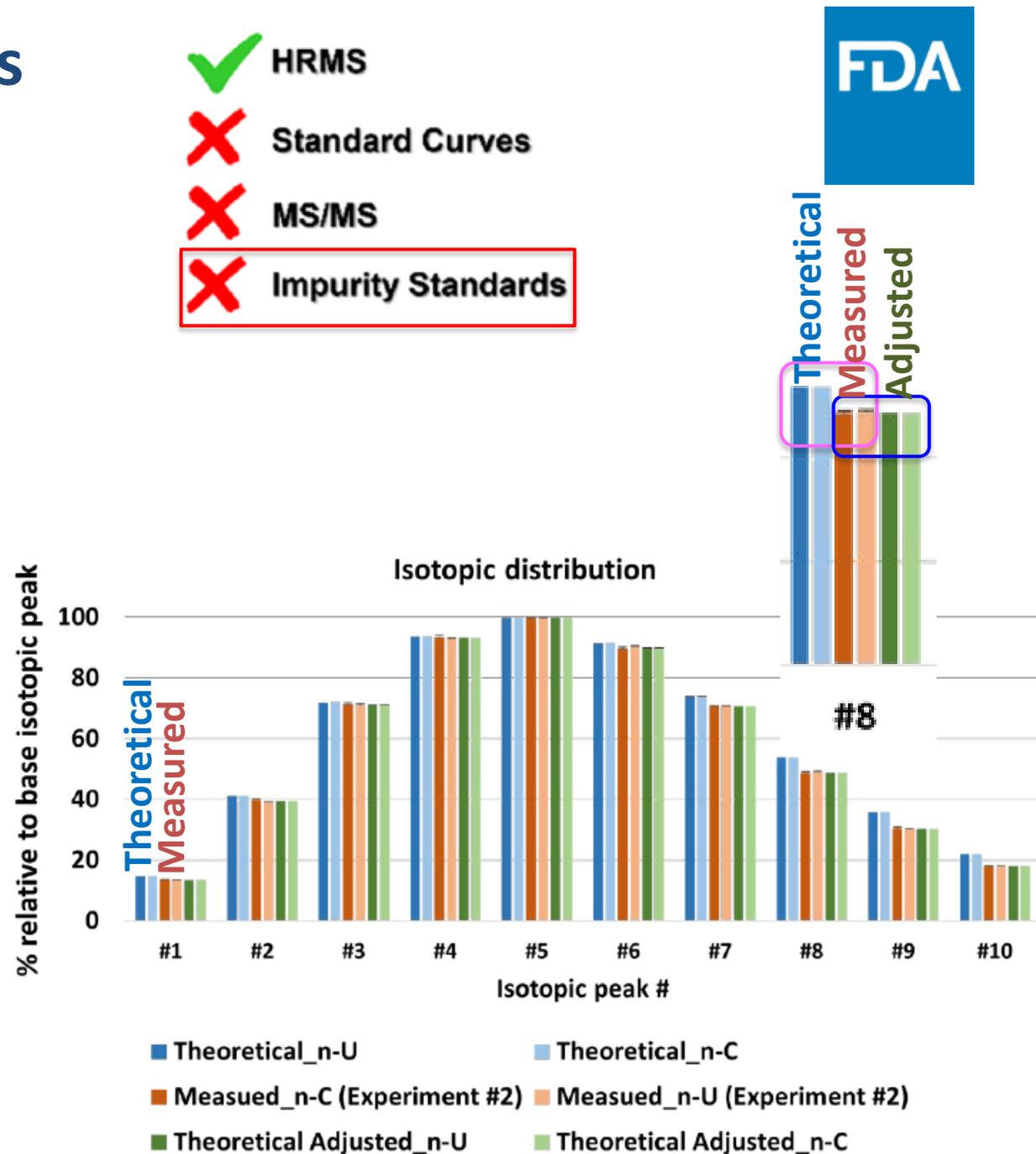


Figure 5. Quantified isobaric compositions of n-U/n-C mixtures using the theoretical isotopic distributions of standards. The same HRMS data as in Figure 2 were reprocessed by replacing the measured isotopic distributions with the theoretical (left panel) or the adjusted theoretical (right panel) distributions of n-U and n-C standards. The quantification results are displayed using the same accuracy plots as denoted in Figure 2A.



? Coeluting isobaric impurity ions

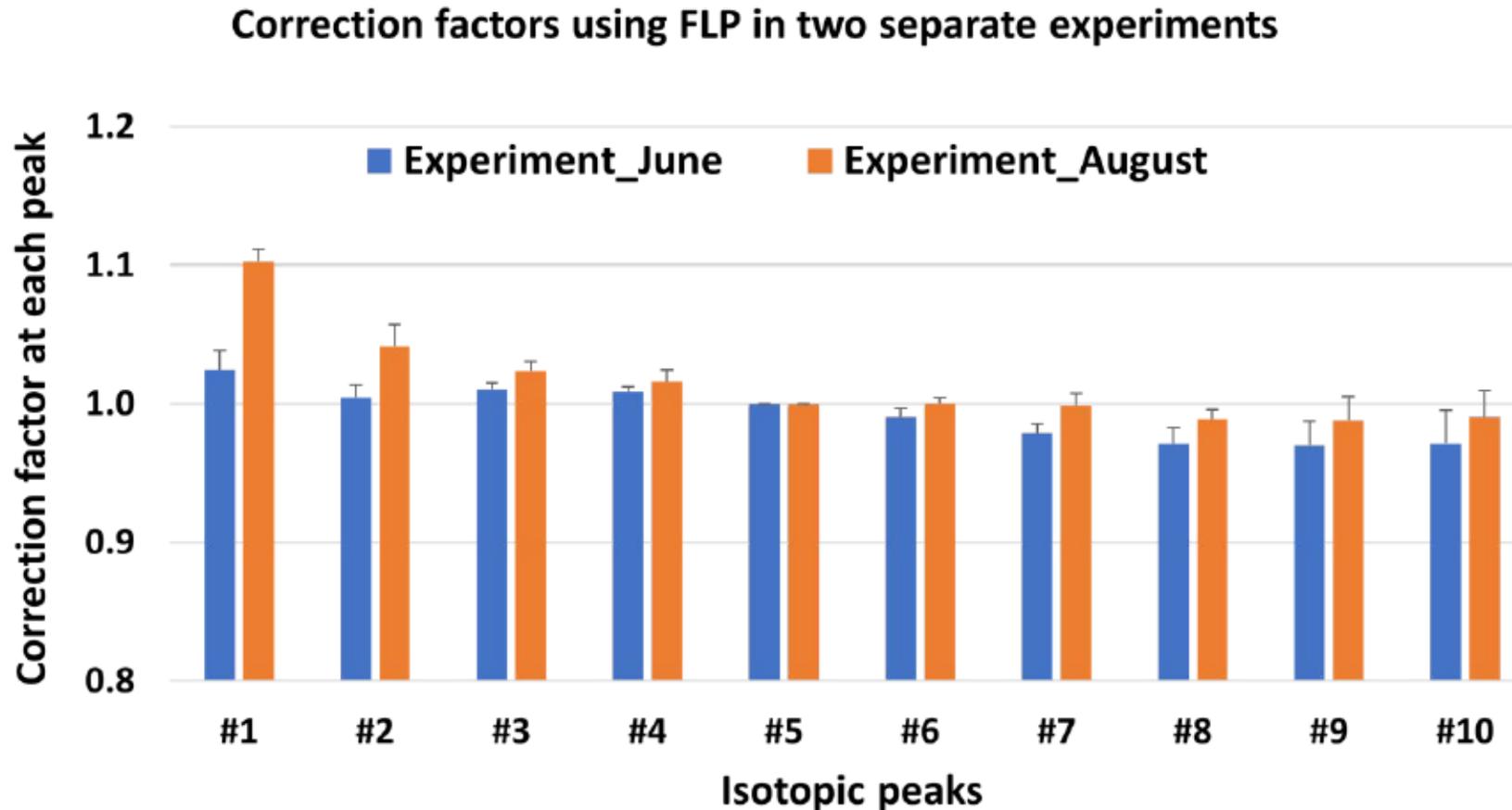


Figure S9. Comparison of correction factors generated by the ratio of the measured vs the theoretical isotopic distributions of FLP from two separate experiments. The measured isotopic distributions were from multiple FLP solution preparations in two separate experiments (Table S4B and Table S4C). The experiment-specific correction factors may be attributed to the instrument's performance while operating under its current tune and calibration settings.

? Coeluting isobaric impurity ions

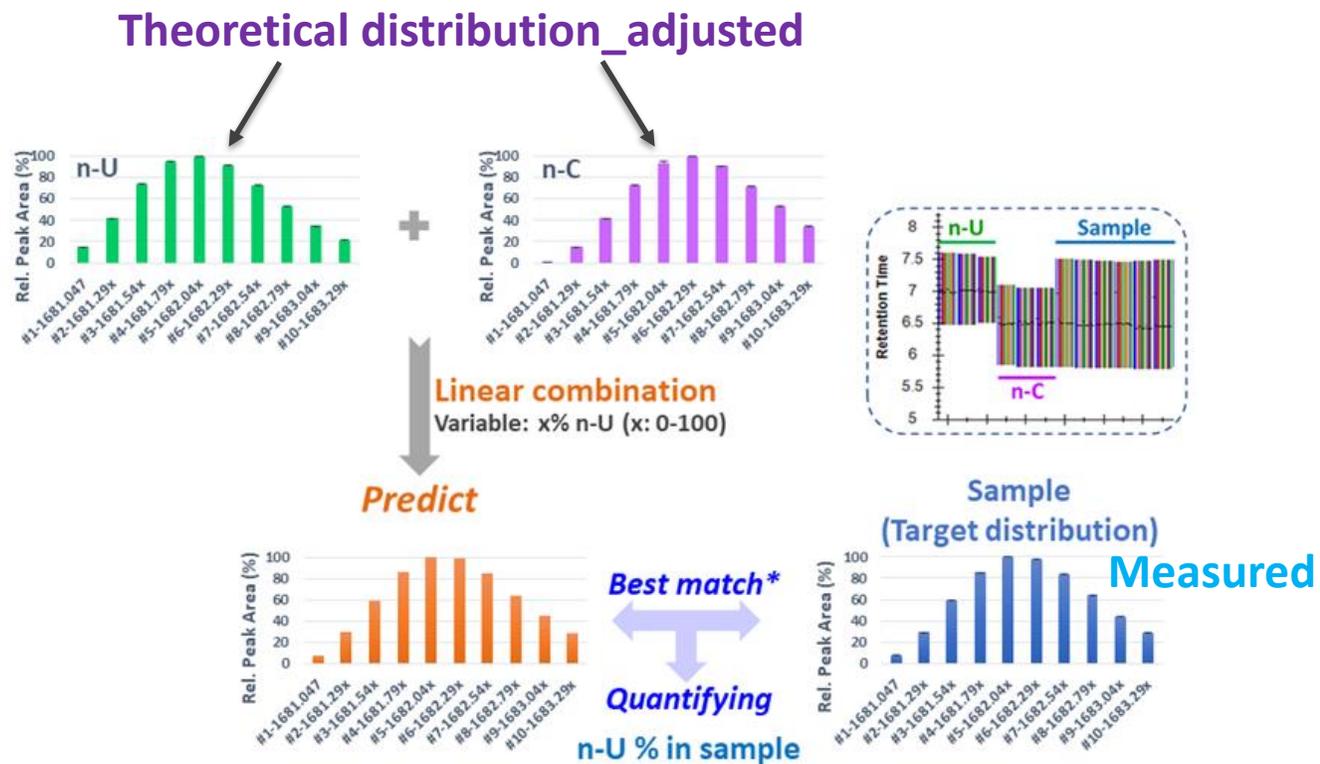
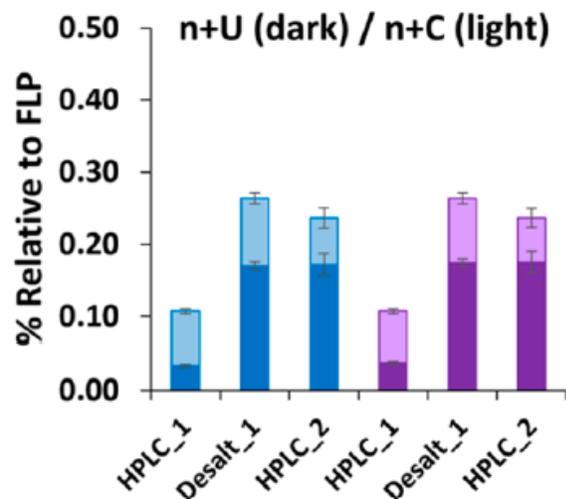
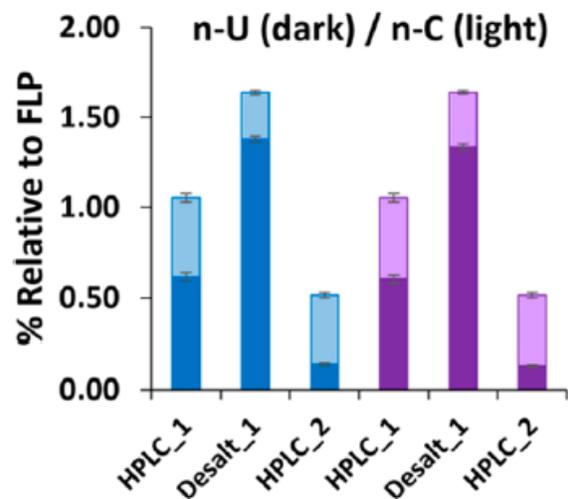
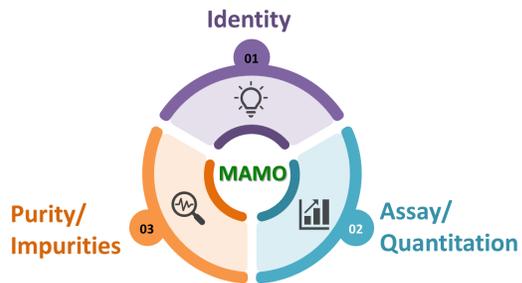


Figure 6. Quantification of U/C deletion and addition impurities using the measured or adjusted theoretical isotopic distributions of n-U/n-C and n+U/n+C. (Left panel) n-U% (dark in color) and n-C% (light). (Right panel) n+U% (dark) and n+C% (light). The y-axis represents quantified impurity amounts relative to FLP. Quantification was performed using the measured (in blue) or adjusted theoretical (in purple) isotopic distribution of n-U/n-C or n+U/n+C. The correction factors used for adjusting the theoretical distributions were generated using the measured vs theoretical isotopic distributions of FLP. Three batches of sample were tested, including two HPLC-purified batches and one desalted batch.

Summary



LC-HRMS-based MAMO platform:

Identity/characterization: RT, m/z,
MS/MS

Quantification: Assay, Purity, Impurities

LC-HRMS method validated for:

Specificity, Linearity, Range, Precision, Accuracy,
LLOQ, Robustness, Excipient



When impurities inseparable by either LC or HRMS:

Coeluting isobaric impurity ion case –
Fully resolved isotopic envelopes enabled by HRMS

Acknowledgement



- **Office of Pharmaceutical Quality Research (OPQR)**

- ✓ OPQR-St. Louis
- ✓ Oligo research team (including **ORISEs***)

* **FDA ORISE Fellowship Program at CDER** through an agreement between the U.S. DOE and FDA

- **OPQ-OGD complex product PSG development Oligo SME triage team**

- ✓ Lead: OPQR
- ✓ Team: OPQ, OGD, OND

- **Office of Generic Drugs (OGD)**

- ✓ ORISE funds
- ✓ ORS – ORISE co-mentoring

- **FDA Research Grants (Critical Path, CSG grant)**

- ✓ PI: OPQR
- ✓ Co-PIs and collaborators: OPQ, OGD, OND, NCTR



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