

USP Symposium, October 7th 2022

Assessment of shelf-life and metabolic viability of a multi-strain synbiotic using standard and innovative enumeration technologies



Dr. Marco Pane

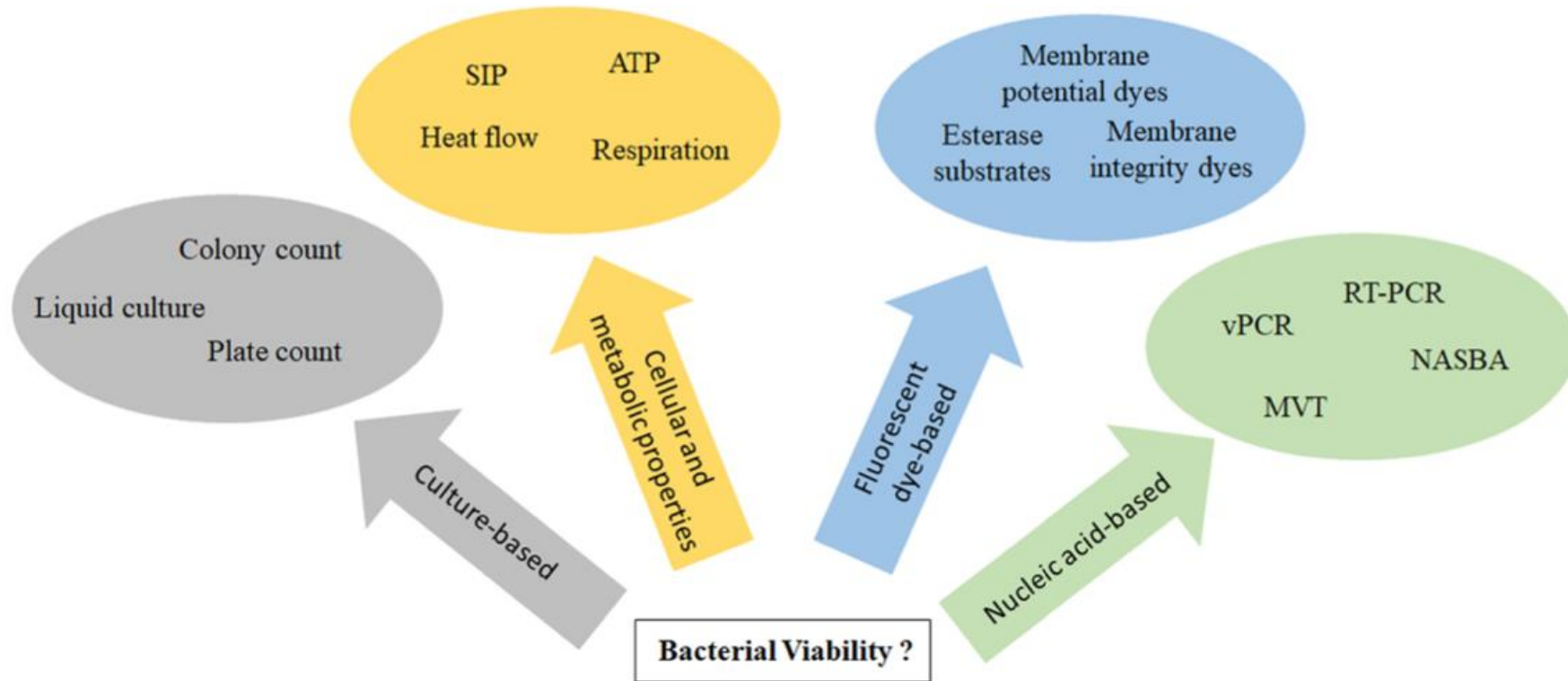


m.pane@probiotical.com

- *R&D Head Probiotical Research*
- *IPA member Scientific and Manufacturing Committee*
- *Board Director Pharmabiotic Research Institute*
- *Expert consultant Probiotic Committee US Pharmacopoeia*
- *Master degree in Food Technology*
- *PhD (ongoing) in Food, Health and Longevity*

Main topics

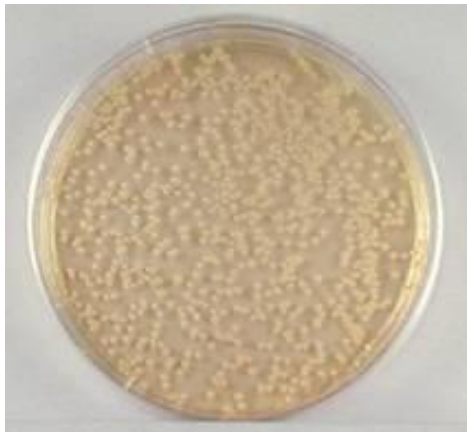
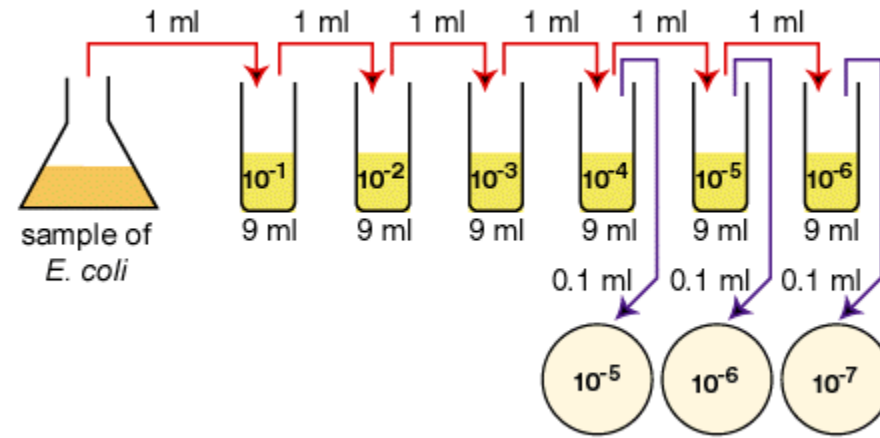
- General principles of Plate Count (PC) and Flow Cytometry (FC) for bacterial enumeration
- Long-term Real-life stability study Probiotic Food Supplement using PC, FC and metabolic assessment
- Case study on VBNC (Viable But Not Cultivable)



Graphical abstract

Representation of various methods for the determination of bacterial viability.

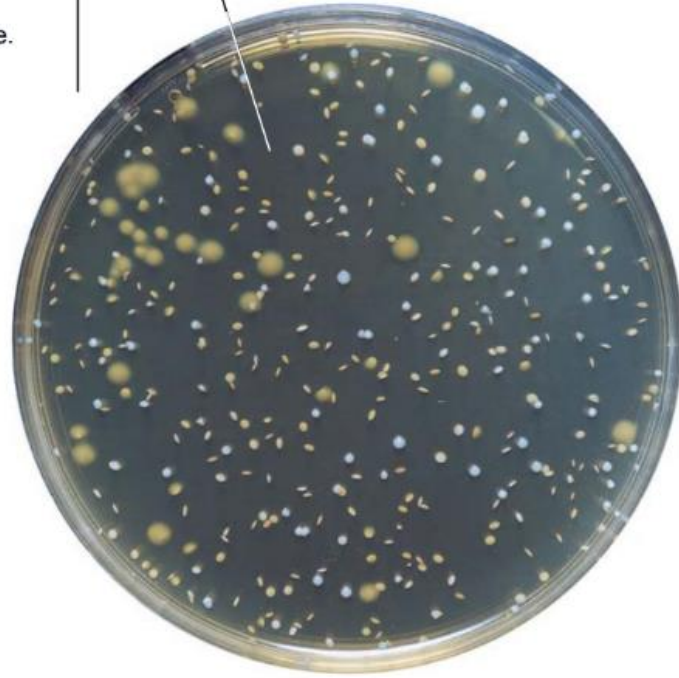
Plate count methods



Lactobacillus casei subsp.
rhamnosus

Typical aspect

White colonies of uniform size.

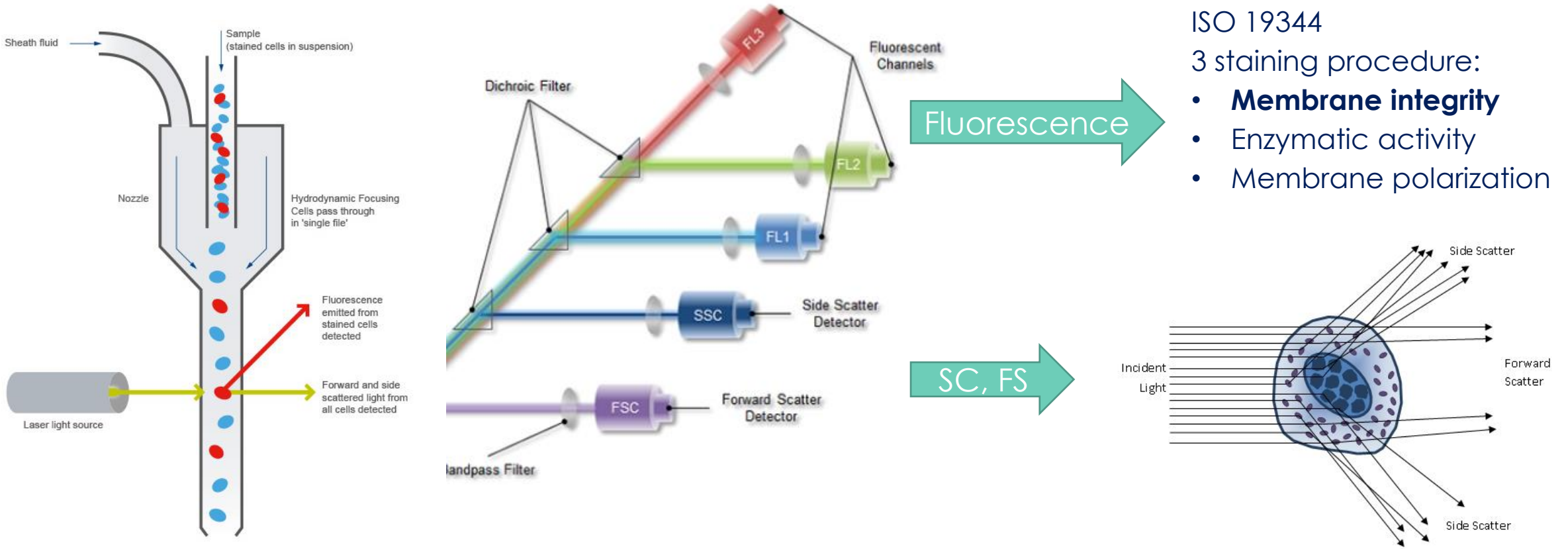


The plate count method is based on the premise that a single bacterium can grow and divide to give an entire colony

Interpretation of plate count methods

Observed result	Usual interpretation	Alternative interpretations
A colony is formed	A viable cell gave rise to the colony	At least one viable cell gave rise to the colony—but it may have been two or more cells coinciding at the same place on the plate or a clump of cells that contained at least one viable individual
No colony is formed	There were no viable cells in the sample	(i) The growth medium and/or incubation conditions were incorrect (ii) The cells were damaged/stressed and therefore unable to grow on solid medium (iii) The population density was low and therefore cell-cell communication could not take place, resulting in no observable growth (iv) Insufficient time was allowed for visible colony development in slowly growing cells

Flow Cytometry (FC) principle



Basically it “just” enumerate cells

Impedance Flow Cytometry

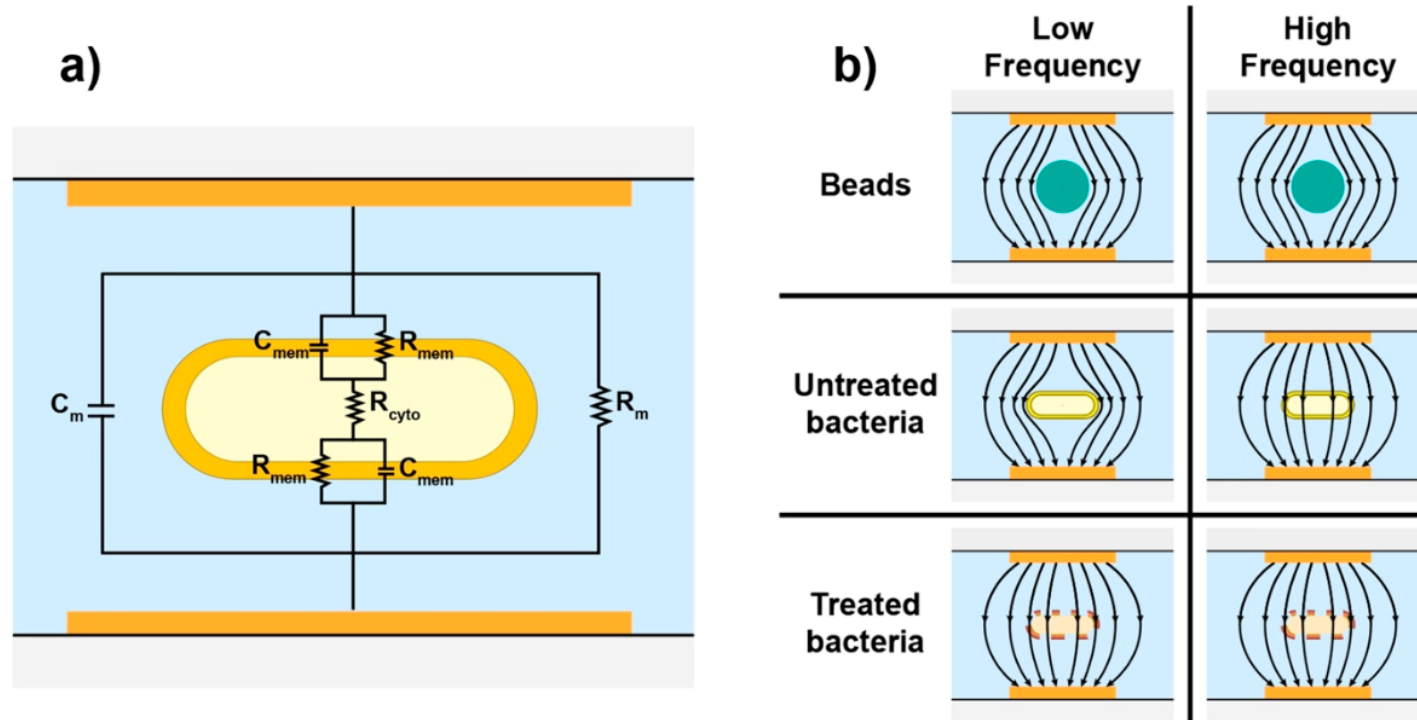


Figure 2. (a) Simplified equivalent circuit model of *E. coli* bacteria. The model is composed of components representing the electrolyte resistance and capacitance (R_m , C_m), the resistance and capacitance of the cell membrane (R_{mem} , C_{mem}) and the resistance of the cell interior (R_{cyto}). (b) Electric field penetration in polystyrene beads and in bacteria with different viability states at low and high frequencies.

INTERNATIONAL STANDARD

**ISO
19344**

**IDF
232**

First edition
2015-12-15

Milk and milk products — Starter cultures, probiotics and fermented products — Quantification of lactic acid bacteria by flow cytometry

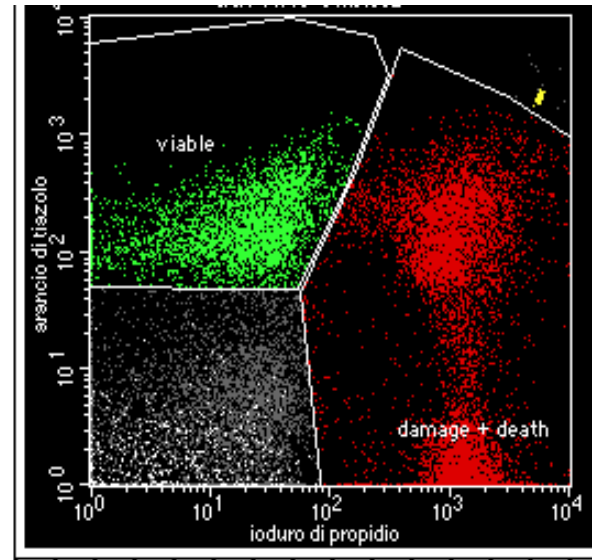
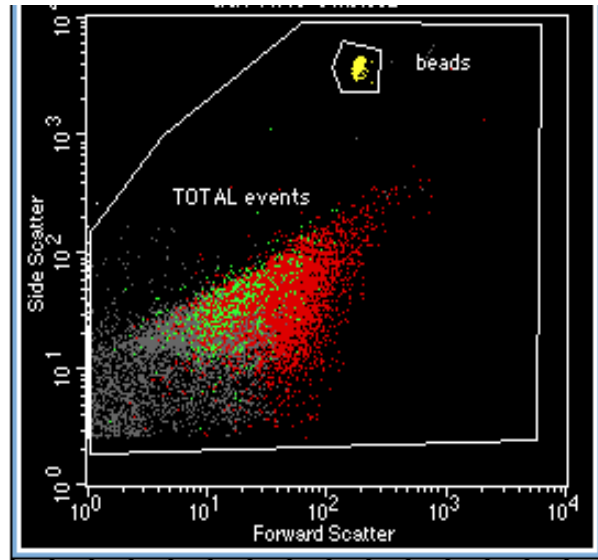
*Lait et produits laitiers — Cultures, probiotiques et produits fermentés
— Quantification de bactéries lactiques par cytométrie en flux*

- **15** labs (5 Countries)
 - **9** different FCM model
 - **10** samples (2 different batch)
 - **8** pure starter culture
 - **1** blend starter culture
 - **1** yoghurt sample
 - **3** staining protocols
 - **2** repetition per lab per strain per protocol
 - **1800** tests
-
- **SAME method for ALL the lactic bacteria!**

Probiotic Enumeration Example: Membrane Integrity

Forward Scatter (FS) =
dimension

Side Scatter (SS) =
granularity and
morphology



Thiazole orange (TO) or equivalent
penetrates all bacteria and stain
the **nucleic acid** with **green
fluorescence** (y axis)

Propidium iodide (PI) penetrates
only bacteria with damaged
membranes with **red fluorescence**
(x axis)

AFU: Active Fluorescent Unit (viable cells)

TFU: Total Fluorescent Unit

(total number of cells: viable + damaged + dead)

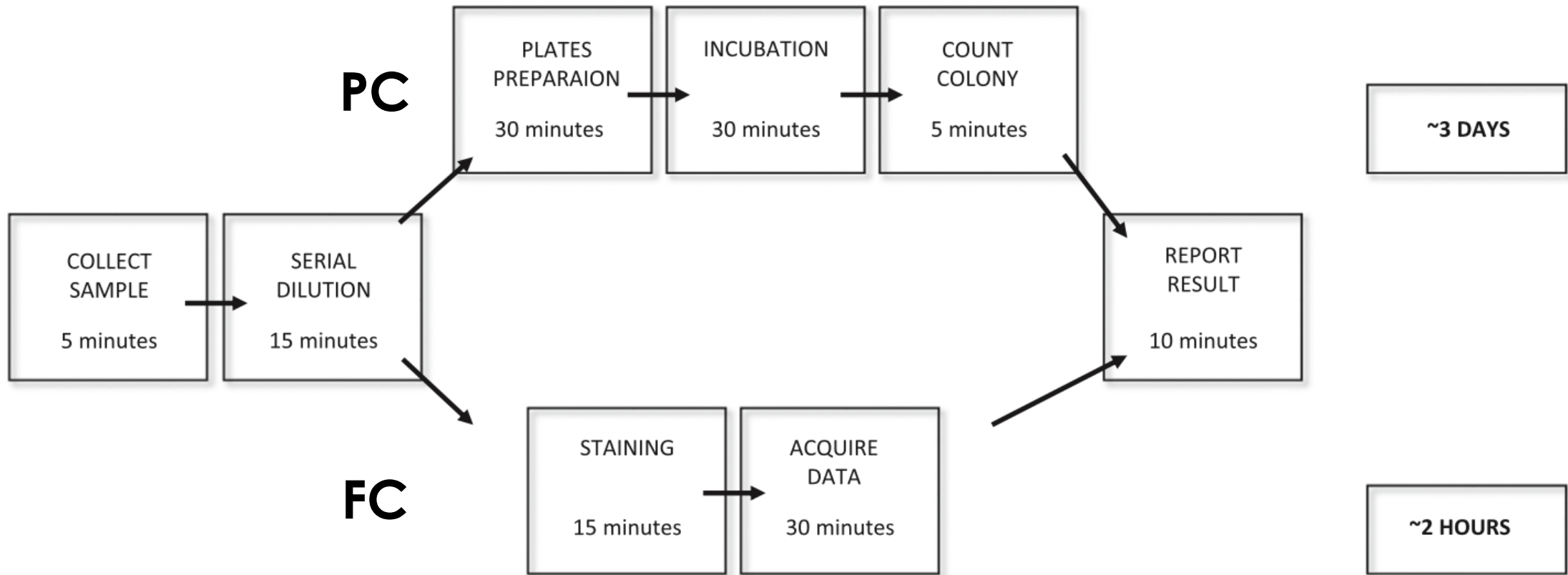


Fig. 1. Work time comparison PC method and FCM method. The plate method requires more time from researcher such as preparation of sterile media, plates and incubation time compared to FCM method which only requires a one-time optimization of the protocol and then only sample staining is required each time.

Bacterial population dynamic during **fermentation**

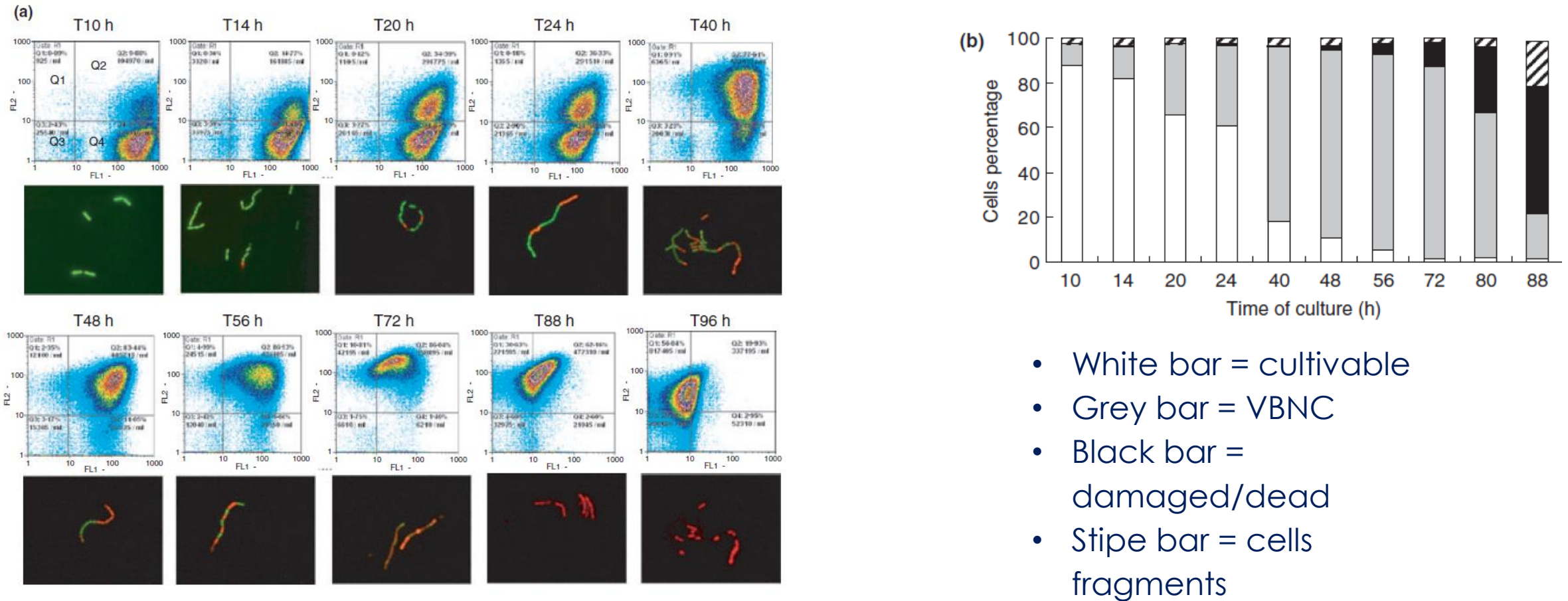


Figure 2 (a) Dot plots and microscopic images of double-stained cells of *Lactococcus lactis* ssp. *cremoris* SK11 with carboxyfluorescein diacetate and propidium iodide dyes during growth. (b) Relative percentage of each population discriminated in dot plots. (□) Green cells; (■) double-stained cells; (■) red cells and (▨) no stained cells.



Contents lists available at [ScienceDirect](#)

Journal of Microbiological Methods

journal homepage: www.elsevier.com/locate/jmicmeth



New insights in enumeration methodologies of probiotic cells in finished products



Chiara Foglia^a, Serena Allesina^b, Angela Amoruso^c, Annachiara De Prisco^c, Marco Pane^{c,*}

^a *Probiotical S.P.A., Enrico Mattei, 3-28100 Novara, Italy*

^b *Biolab Research S.r.l., Enrico Mattei, 3-28100 Novara, Italy*

^c *Probiotical Research S.r.l., Enrico Mattei, 3-28100 Novara, Italy*

Bacterial heterogeneity and population dynamic during product **STABILITY:**
the use of predictive microbiology

Table 1. Storage conditions according to International Conference on Harmonization (ICH) pharma guidelines Q1A for the development of new drug products. RH: Relative Humidity.

Study	Storage Condition	Minimum time needed for data submission	Control points (months)
Refrigerated condition ^b	5°C ± 3°C	12 ^b -24 ^a months	0, 3, 6, 12, 18, 24
Long Term ^{a-b}	Zone II 25°C ± 2°C 60% ± 5% RH Zone IVb 30°C ± 2°C/ 75% ± 5% RH	12 ^b -24 ^a months	0, 3, 6, 12, 18, 24
Accelerated ^{a-b}	40°C ± 2°C 75% ± 5% RH	12 ^b -24 ^a months	0, 1, 2, 3, 6, 18, 24

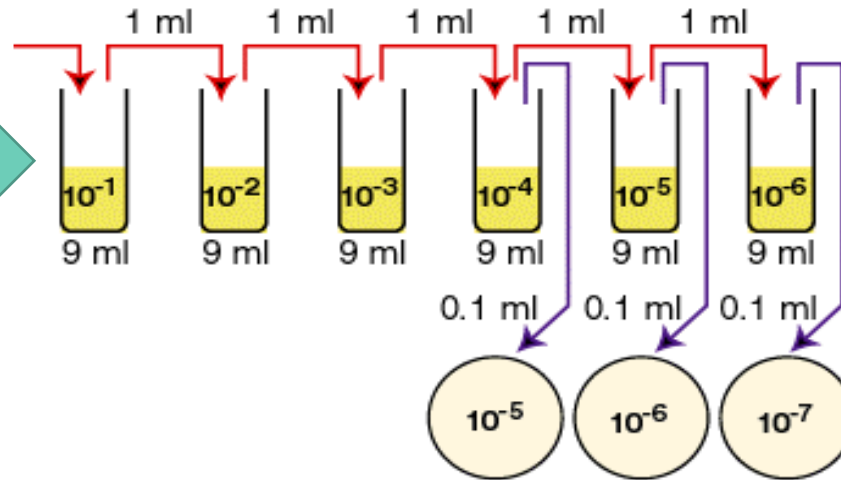
a. First experiment (CFU vs AFU)

b. Second experiment (CFV vs AFU vs TFU vs pH)

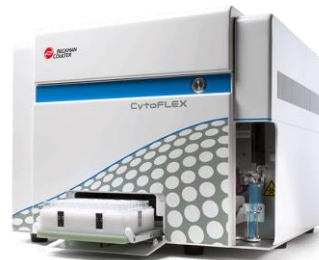
a) Measurand: cryoprotected freeze-dried single strain probiotic (*L. rhamnosus*) supported in maltodextrin

3 FREEZE DRYED INDUSTRIAL BATCH PRODUCTS

Enumeration



CFU: Colony Forming Unit



AFU: Active Fluorescent Unit Membrane Integrity
TFU: Total Fluorescent Unit

Long-term stability with PC and FC

Fig. 1. Effect of storage condition at 25 °C on bacterial stability. Plot of $\ln(N_t/N_0)$ versus time (mean of 3 values) (dotted line: FC, straight line PC).

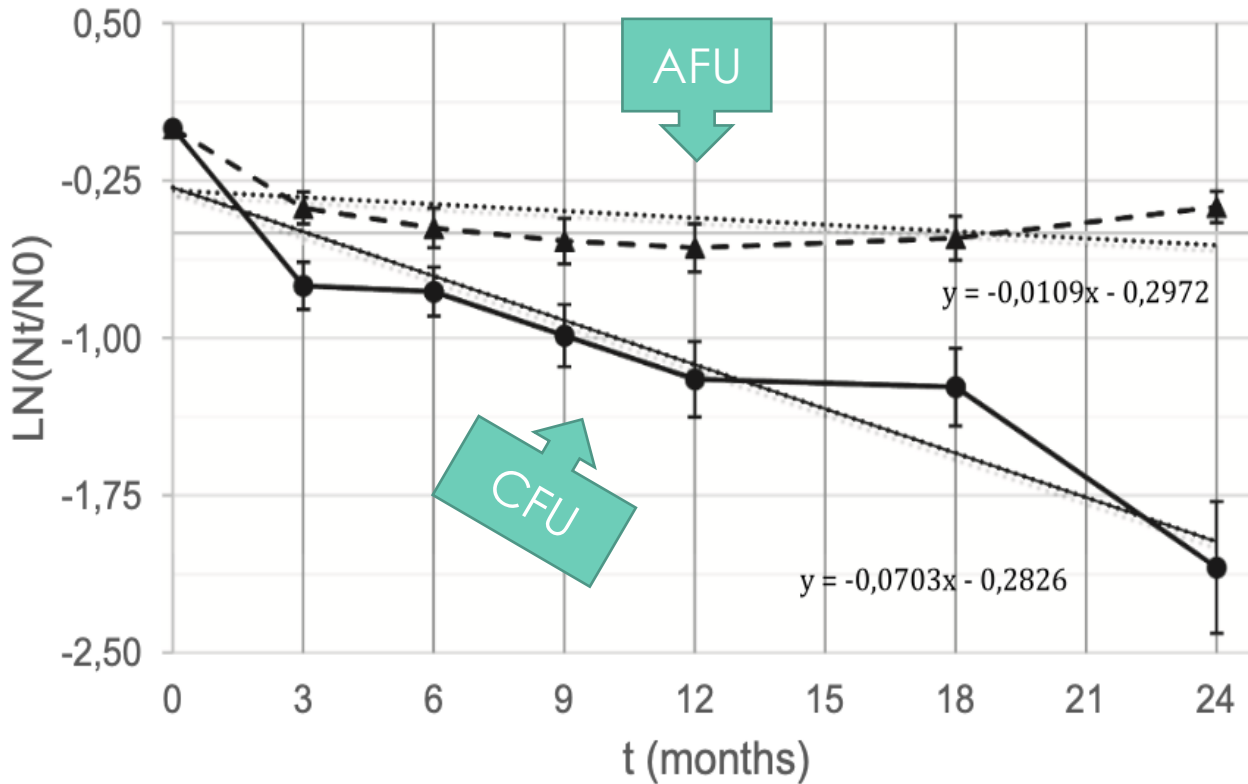


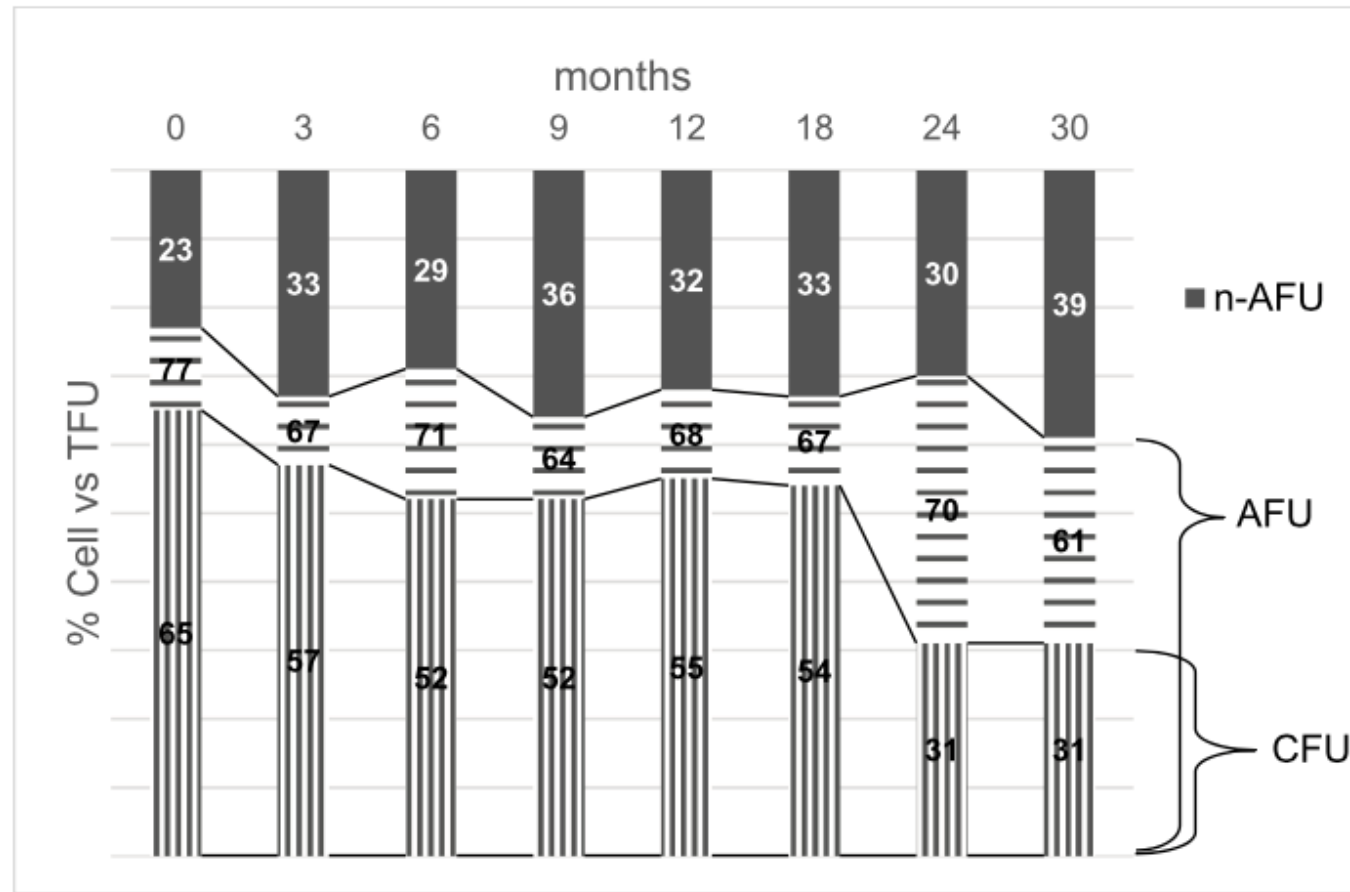
Table 3

Cultivability and membrane integrity destruction rate (k) and decimal reduction time (D_1) of probiotic samples at different temperatures.

Temperature °C (T K)	Destruction rate k (months ⁻¹)		k_{PC}/k_{FC}	Decimal reduction time D_1 (months)	
	PC	FC		PC	FC
25 °C (298,15)	0.0386	0.0049	7.877	59.7	469.9
30 °C (303,15)	0.0703	0.0109	6.449	32.8	211.2
40 °C (313,15)	0.7098	0.1033	6.871	3.2	22.3



doi: 10.1016/j.mimet.2020.105993.

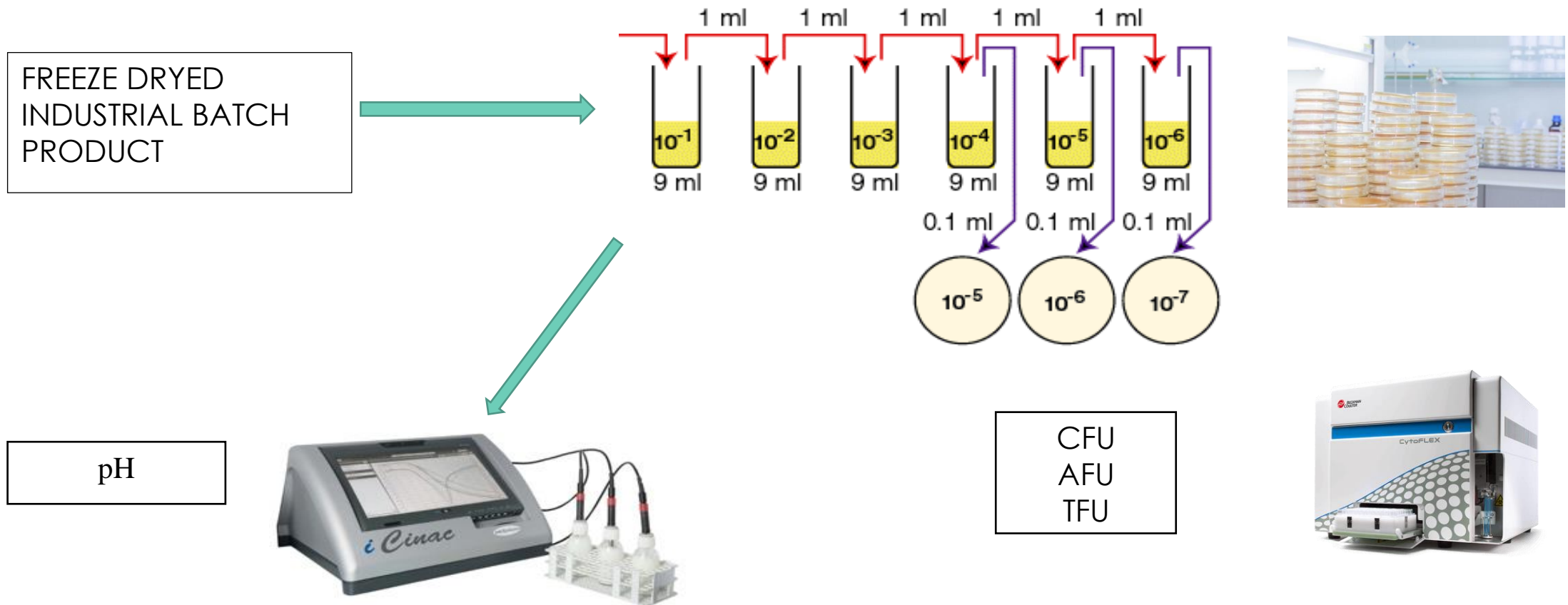


AFU-CFU = VBNC ?

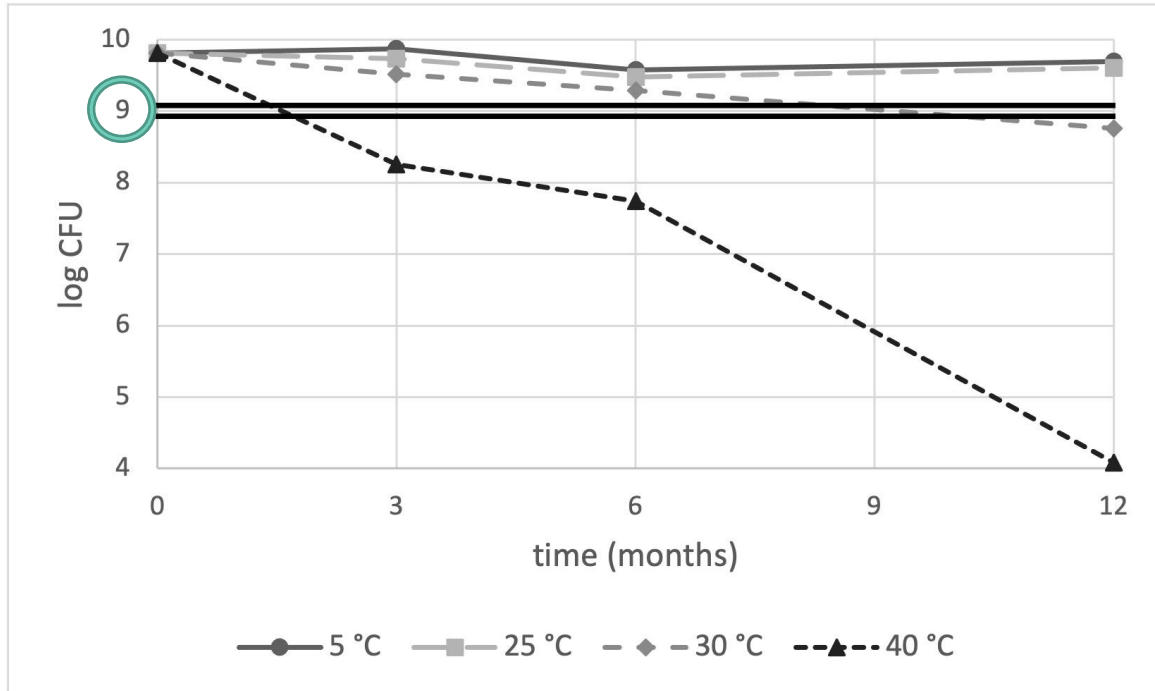
Fig. 5. Bacterial heterogeneity distribution over time at 25 °C expressed as the % of cells (CFU, AFU and n-AFU) vs TFU (100% of the population at each time point). Relative % of AFU is the sum of AFU + CFU values; for example, t0 is represented by 65% CFU vs TFU, 77% AFU vs TFU and 23 % n-AFU vs TFU). First line from the top is representative of the AFU trend vs time. Second line from the top is representative of CFU trends vs time.

doi: 10.1016/j.mimet.2020.105993.

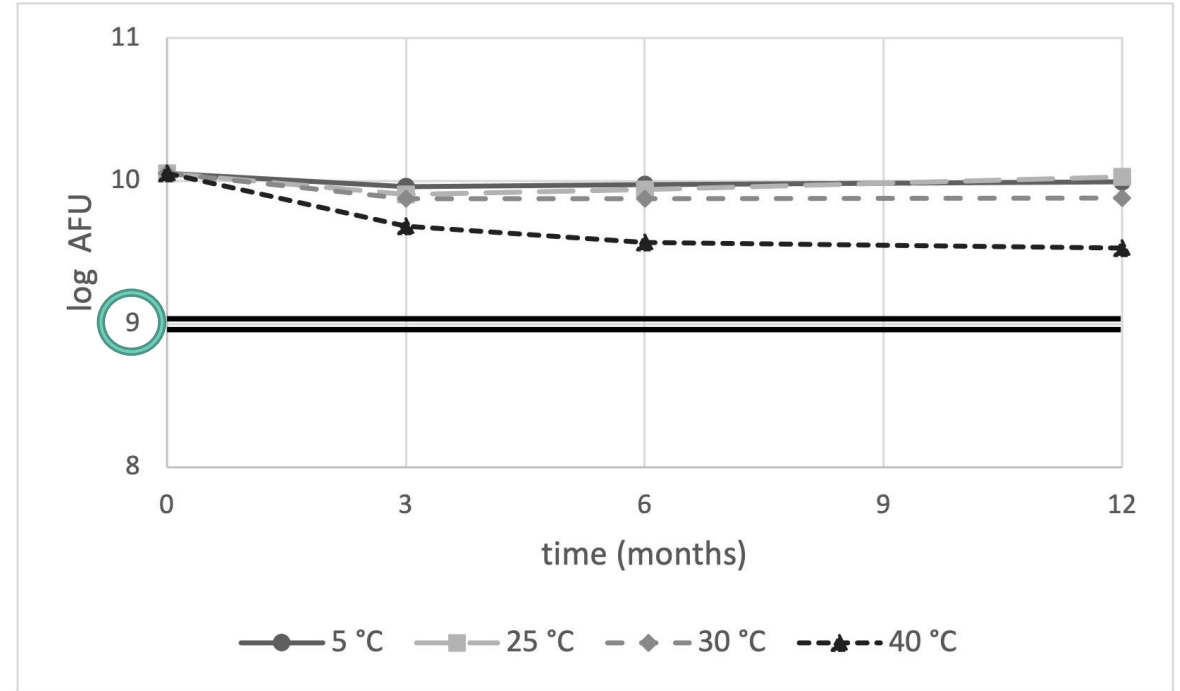
b) Measurand: cryoprotected freeze-dried multi-strain probiotic supported in inulin (accepted for publication 27/09/22)



CFU and AFU trends at 5, 25, 30 and 40 °C for 12 months



CFU



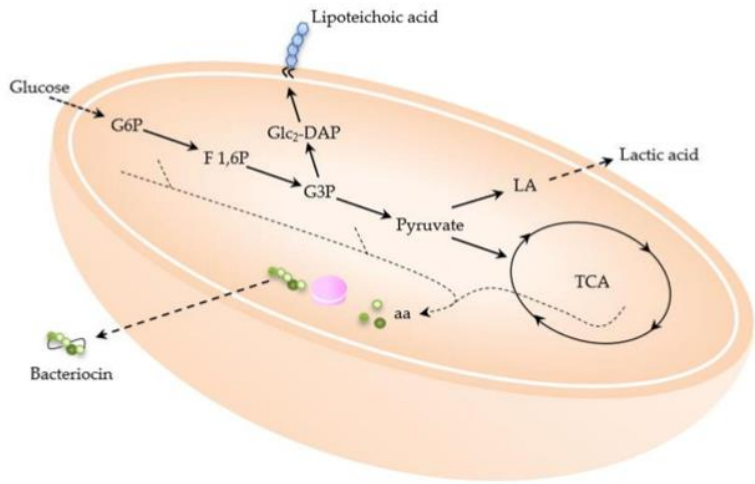
AFU

Destruction rate and Decimal Reduction Time

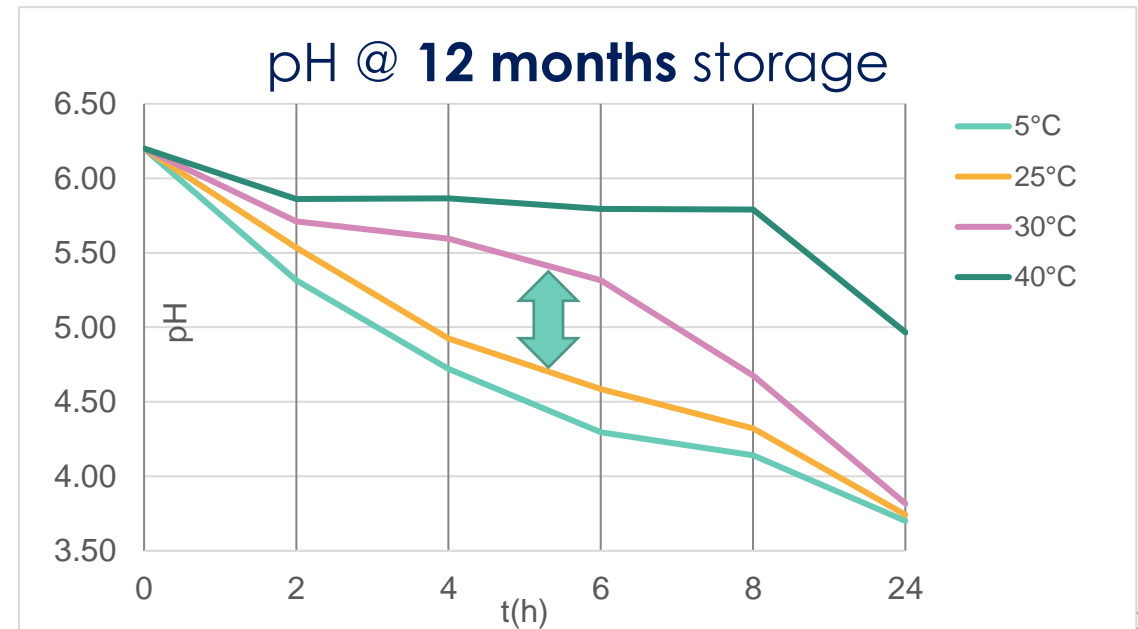
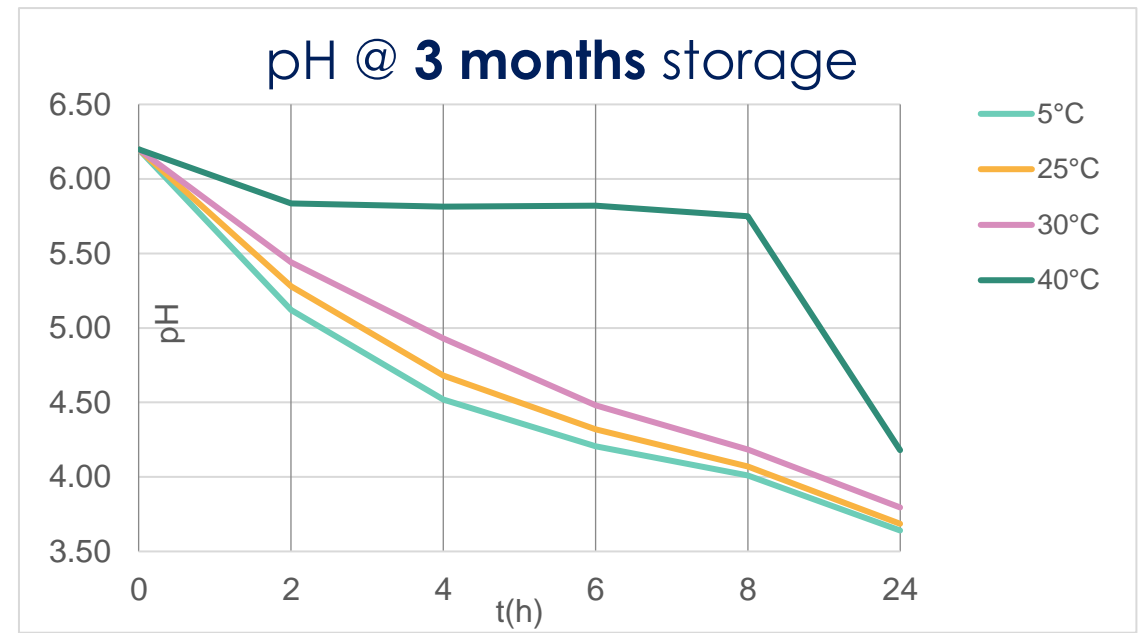
Table 2. Bacterial/probiotic cultivability and membrane integrity destruction rate (k) and decimal reduction time (D₁) of synbiotic samples at different temperatures.

Temperature	Destruction rate k (months ⁻¹)			Decimal reduction time D ₁ (months)		
	CFU	AFU	TFU	CFU	AFU	TFU
5 °C (278.15 °K)	0.0247	0.0025	0.0020	93.2	921.0	1151.3
25 °C (298.15 °K)	0.0468	0.0034	0.0025	49.2	677.2	932.0
30 °C (303.15 °K)	0.1844	0.0161	0.0141	12.5	143.0	163.3
40 °C (313.15 °K)	0.6983	0.1417	0.1212	3.3	16.2	19.0

CFU= Colony Forming Units; AFU= Active Fluorescent Units; TFU= Total Fluorescent Units

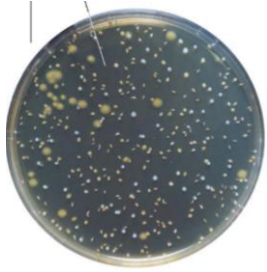


**pH as a metabolism proxy
thanks to lactic acid
production**

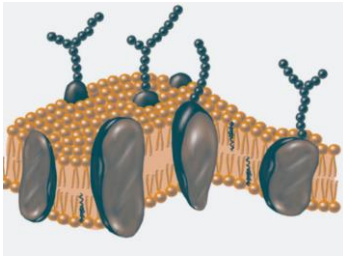


Decay Kinetics of TFU, AFU, CFU and pH @12 months

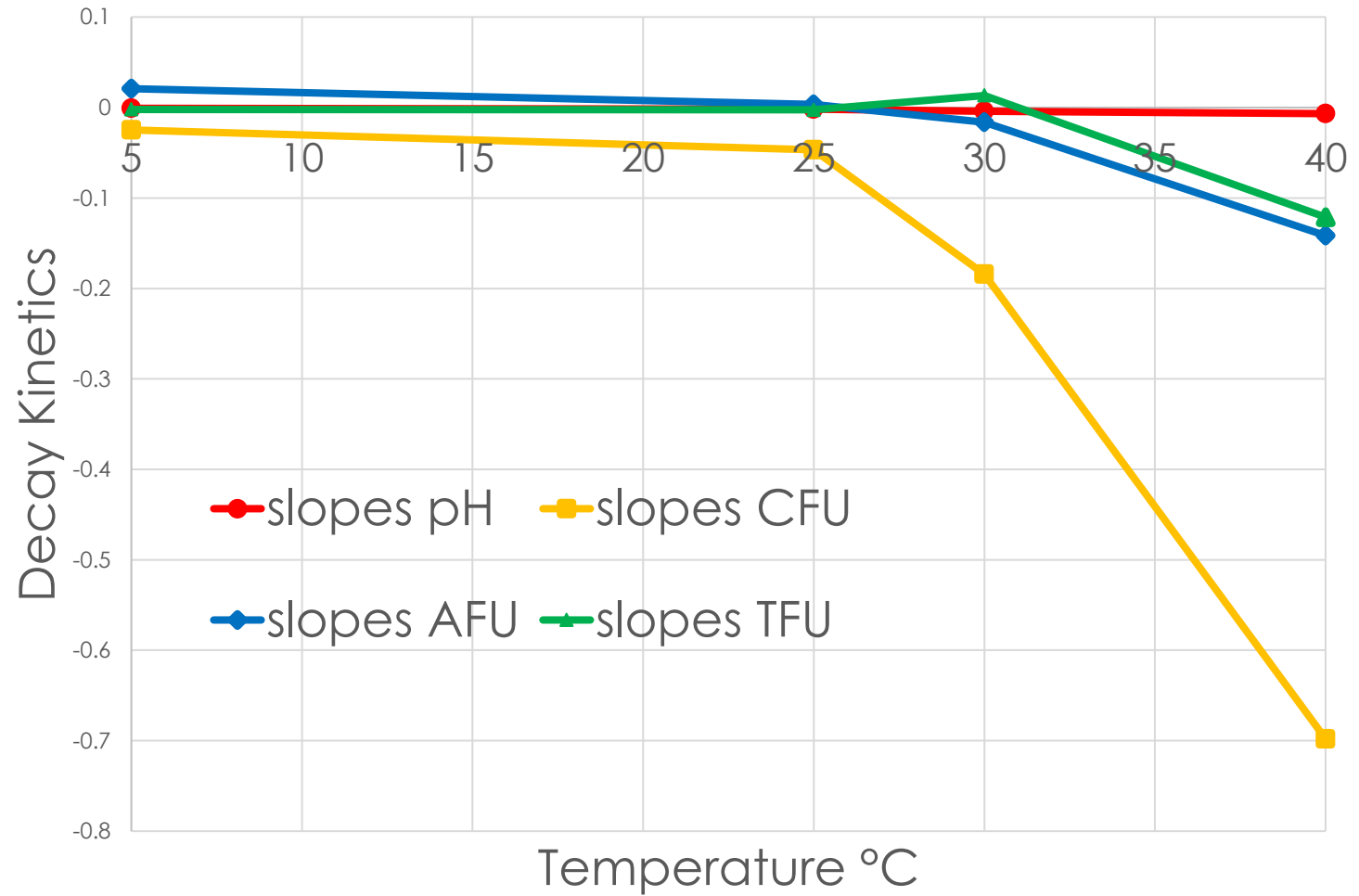
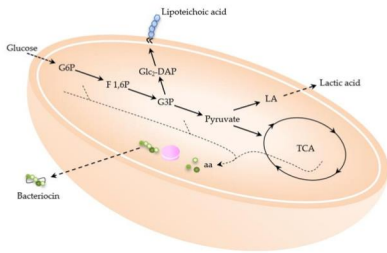
CFU



AFU
TFU



pH



Viable But Non-Cultivable (VBNC) cells

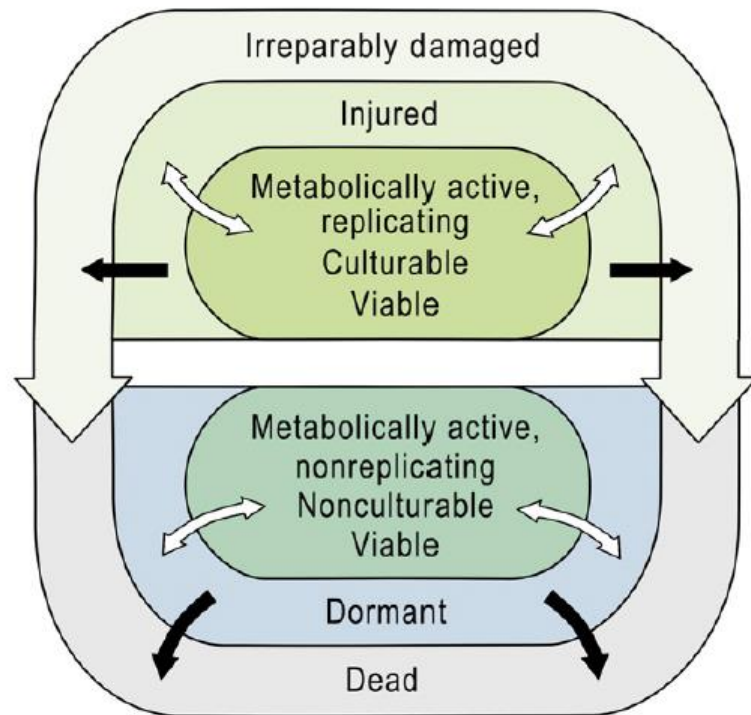


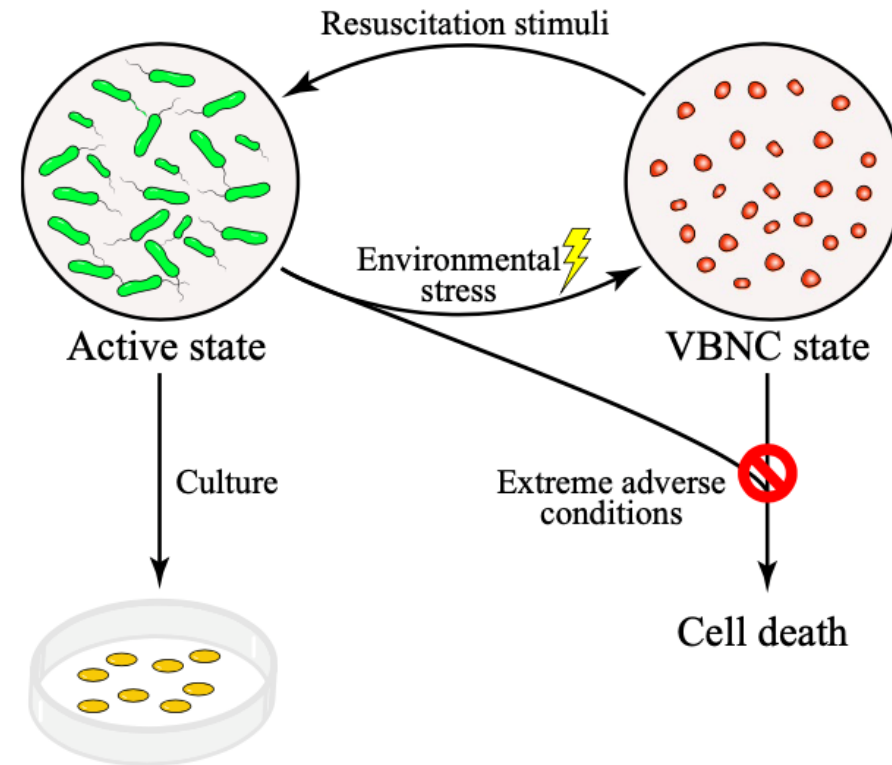
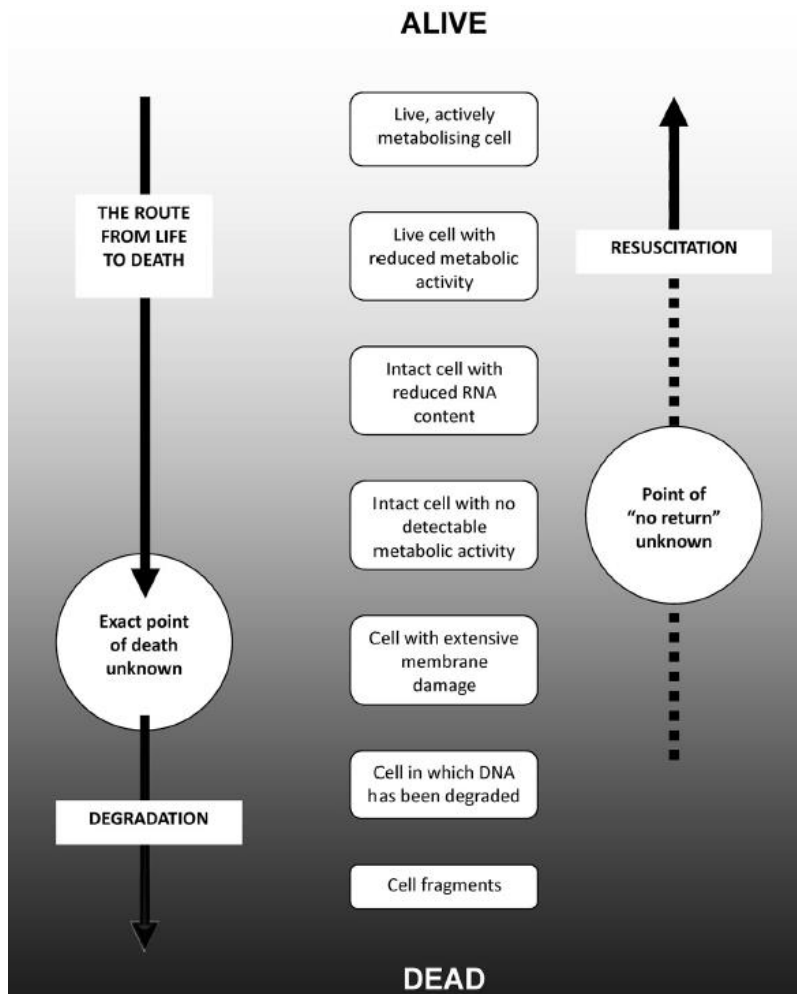
Fig. 1. A concept map for probiotic strains that describes metabolically active, replicating/culturable/viable states and the transitions that are possible. The arrow on the perimeter and the black one-way arrows indicate that once a cell is non-viable/dead it does not return to a viable state.

“Microbes exists in a variety of growth phases and metabolic states depending on environmental conditions and stressor, and **only a subset of these involve active replication.** The convention that viable microbes must be capable of forming colonies excludes not only dead or irreparably damaged organisms but also live microbes that have adapted to environmental stress by becoming dormant (VBNC state).”

from Davis, 2014

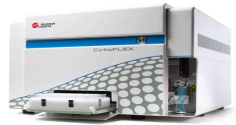
<http://dx.doi.org/10.1016/j.mimet.2014.04.012>

Is it possible to “see” VBNC and prove that they could revert to CFU (close the gap between AFU and CFU)?

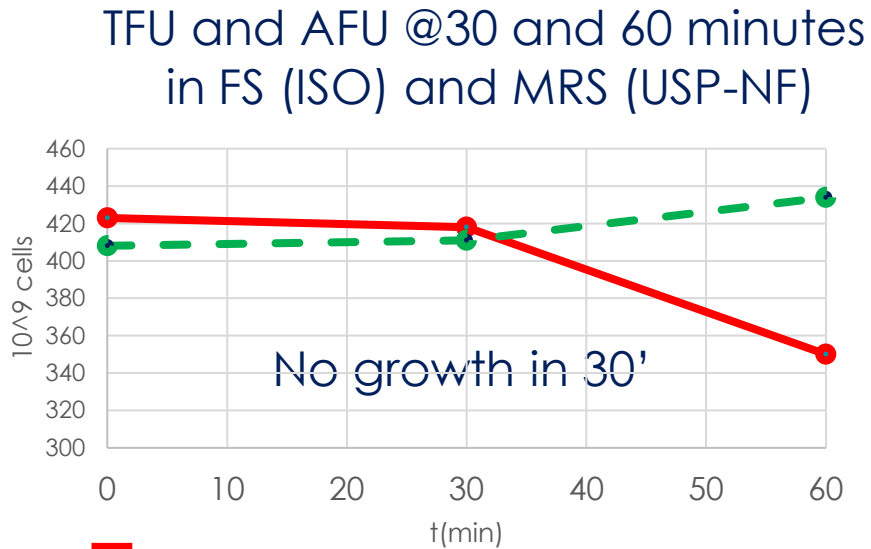


<https://doi.org/10.1007/s42995-020-00041-3>

c) Measurand: 400×10^9 AFU/g cryoprotected freeze-dried *Lactobacillus acidophilus* supported in maltodextrin



● SF AFU
● MRS AFU



Vs AFU 82%

USP vs ISO +33%

Vs AFU 57%

330×10^9 CFU

230×10^9 CFU

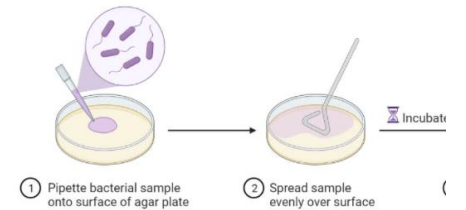
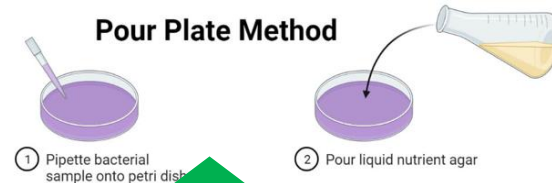
10 replica in triplicate later ..

USP

ISO

Spread Plate Method

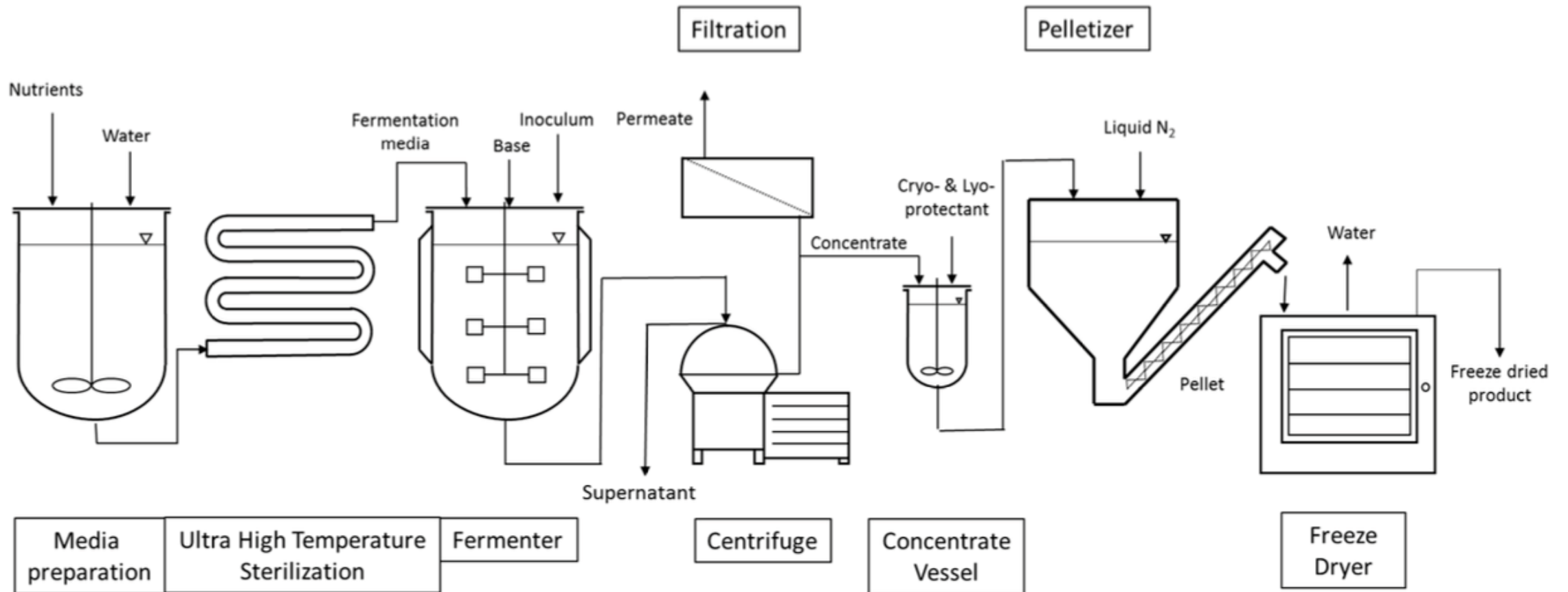
Pour Plate Method



USP-NF: 30 min incubation

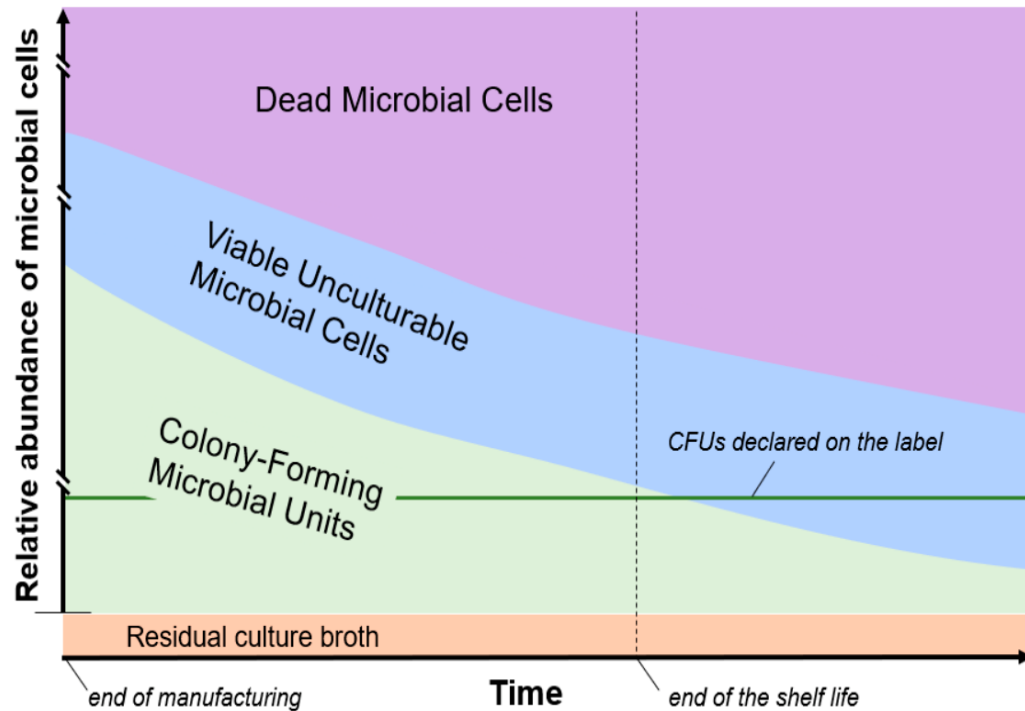
ISO 20128: 0 min incubation

★ The industrial production of probiotics is stressful for the bacterial cell



doi:10.3390/microorganisms7030083

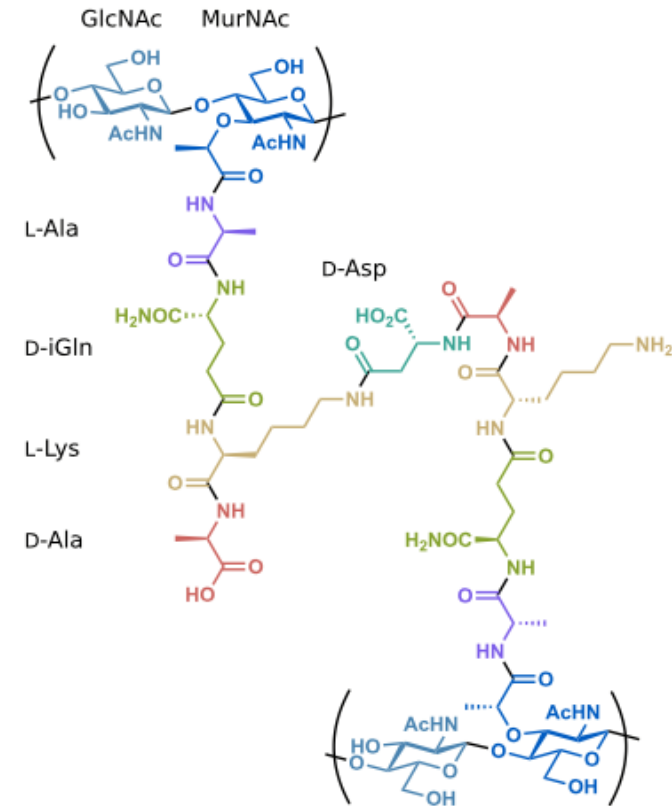
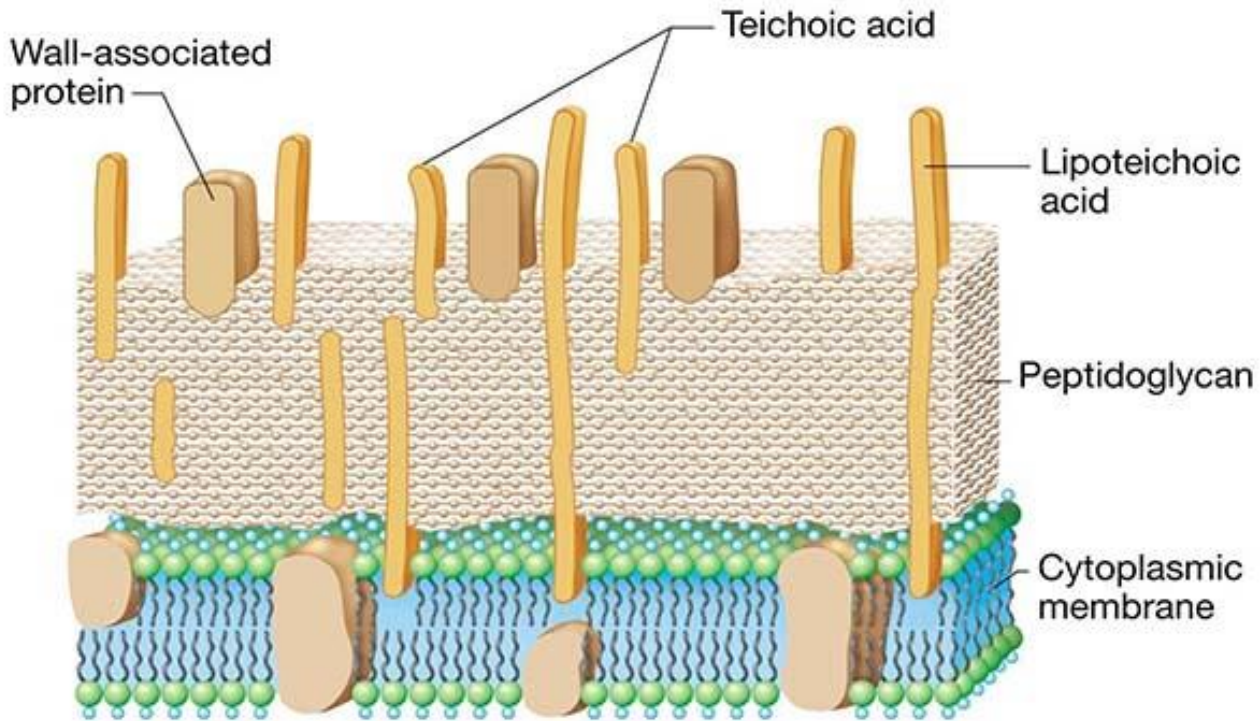
★ Generally probiotic food supplements are characterized by a heterogeneous bacterial population



Can we expect that some biological function are better represented by TFU and AFU than CFU ?

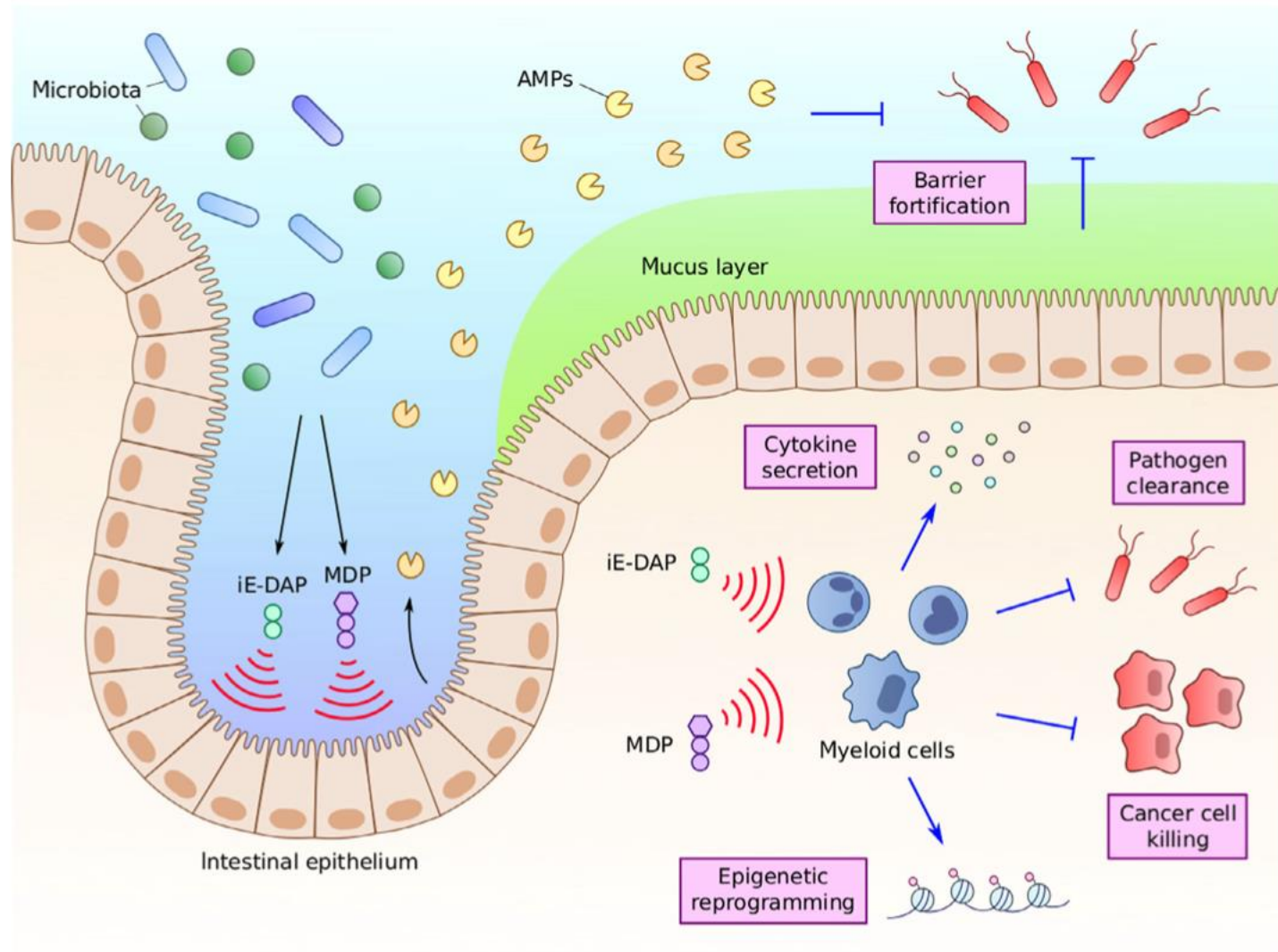


AFU and TFU are representative of cells number and ... their cell wall.



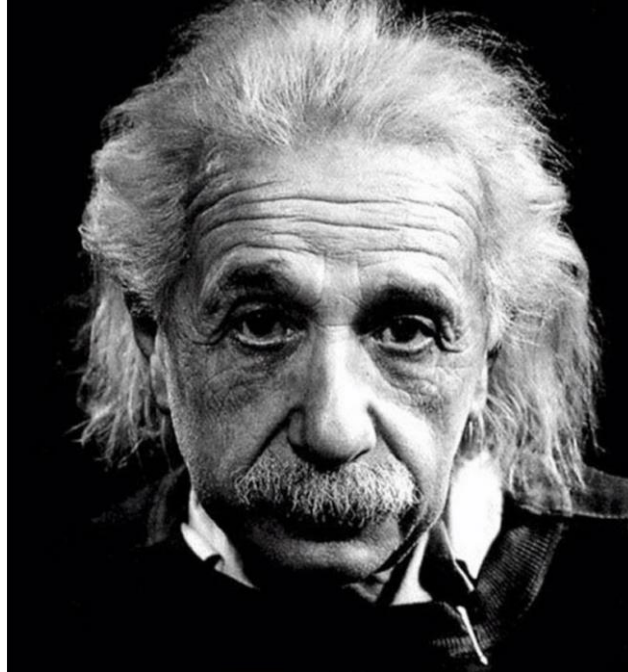
doi: 10.1002/cti2.1095

★ Peptidoglycan as immune-modulators



CFU could underestimate the effective quantity of functional probiotic cells

doi: 10.1002/cti2.1095



“It is the theory (method) which decides what can be observed”



probiotical.com